



Scaling from Traits to Ecosystems: Developing a General Trait Driver Theory via Integrating Trait-Based and Metabolic Scaling Theories

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Abstract

Aim: More powerful tests of biodiversity theories need to move beyond species richness and explicitly focus on mechanisms generating diversity via trait composition. The rise of trait-based ecology has led to an increased focus on the distribution and dynamics of traits across broad geographic and climatic gradients and how these distributions influence ecosystem function. However, a general theory of trait-based ecology, that can apply across different scales (e.g. species that differ in size) and gradients (e.g. temperature), has yet to be formulated. While research focused on metabolic and allometric scaling theory provides the basis for such a theory, it does not explicitly account for differences in traits within and across taxa, such as variation in the optimal temperature for growth. Here we synthesize trait-based and metabolic scaling approaches into a framework that we term 'Trait Driver Theory' or TDT. It shows that the shape and dynamics of trait and size distributions can be linked to fundamental drivers of community assembly and how the community will respond to future drivers. To assess predictions and assumptions of TDT, we review several theoretical studies and recent empirical studies spanning local and biogeographic gradients. Further, we analyze how the shift in trait distributions influences ecosystem processes across an elevational gradient and a 140-year-long ecological experiment. We show that TDT provides a baseline for (i) recasting the predictions of ecological theories based on species richness in terms of the shape of trait distributions and (ii) integrating how specific traits, including body size, and functional diversity then 'scale up' to influence ecosystem functioning and the dynamics of species assemblages across climate gradients. Further, TDT offers a novel framework to integrate trait, metabolic/allometric, and species-richness-based approaches to better predict functional biogeography and how assemblages of species have and may respond to climate change.



1. INTRODUCTION

Understanding and explaining species richness patterns have had far-reaching influence on the development of ecology. Biodiversity science strives to understand the drivers and consequences of variation in the number of species and how species abundances change across spatial and temporal scales (MacArthur, 1972; Rosenzweig, 1995). These changes in species richness have also been linked with changes in ecosystem functioning. The Biodiversity–Ecosystem Functioning (BEF) hypothesis states that ecosystems with greater biodiversity are more productive and stable (Naeem et al., 1994; Tilman, 2001; Tilman et al., 1997). Attempts to answer these questions have led to debates that polarized the field (Wardle, 2002), and a growing consensus that species numbers alone do not inform us about all important aspects of ecosystem functioning and community responses to environmental change (Chapin et al., 2000; Díaz and Cabido, 2001; Diaz et al., 2007; Stevens et al., 2003).

More recently, trait-based approaches have focused on recasting classical questions from the species richness literature (Hillebrand and Matthiessen, 2009; Lamanna et al., 2014; Lavorel and Garnier, 2002; McGill et al., 2006; Violle et al., 2007). Instead of species richness, there is an attempt to focus on functional traits and diversity in trait values (Díaz and Cabido, 2001; Lavorel and Garnier, 2002; Mason et al., 2005; Petchey and Gaston, 2002; Roscher et al., 2012). In addition, metabolic scaling theory or MST has focused on the central role of body size as a critical driver of ecological, ecosystem, and evolutionary patterns and processes (Enquist et al., 1998, 2003; Gillooly et al., 2005; Savage et al., 2004). One could also ask about diversity in the number and/or range of trait or body size values, and to some degree, this depends on how traits are defined. As discussed by Dell et al. (2015) and Pawar (2015) in this issue, the premise is that measures of traits, including body size, can better reveal the mechanisms and forces that ultimately structure biological diversity (Grime, 2006; McGill et al., 2006; Stegen et al., 2009) and increase the generality and predictability of ecological models (Díaz et al., 2004; Kattge et al., 2011; Webb et al., 2010). Trait-based approaches have especially received attention for plant life histories and strategies due to a renewed interest in measuring traits across different environments and scales (Craine, 2009). While this has long been part of comparative physiology and ecology (see Arnold, 1983; Grime, 1977), it is now being heralded as its central paradigm (Craine, 2009;

Westoby and Wright, 2006). Similarly, trait-based approaches are being used to disentangle the forces that structure larger scale biodiversity gradients (Belmaker and Jetz, 2013; Han et al., 2005; Reich, 2005; Reich and Oleksyn, 2004; Safi et al., 2011; Swenson and Enquist, 2007) and to predict large-scale ecosystem shifts due to climate change (Elser et al., 2010; Frenne et al., 2013).

1.1 Central Limitations of Trait-Based Ecology

An important limitation to developing a more predictive trait-based ecology is that its focus and implementation have relied almost entirely on empirical correlations and null models (for example, see discussion in Swenson, 2013). There is a need for theory and quantitative arguments to move beyond pattern searching. Further, trait-based ecology has largely developed independently from MST, where the role of body size—arguably a key trait—is central to scaling up organismal processes. Nonetheless, a key focus of trait-based ecology is to identify the general processes underlying trait-based ecology (Enquist, 2010; Shipley, 2010; Suding et al., 2008b; Webb et al., 2010; Weiher et al., 2011). Such an advance would help guide the explosion of trait-based data collection (Dell et al., 2013; Kattge et al., 2011), develop a more predictive ecology, and organize rapidly developing directions in trait-based ecology (Boulangeat et al., 2012; Funk et al., 2008; Lavorel et al., 2011; McGill et al., 2006; Shipley, 2010; Suding et al., 2008b).

Another limitation is the debate about whether biodiversity, trait diversity, or both are important for ecosystem functioning (Hooper et al., 2004; Loreau et al., 2001). We agree with Cardinale et al. (2007) that this debate is largely a false dichotomy. Increasingly, the evidence shows that both the number of species and types of species in an ecosystem impact biomass production. For example, focusing solely on species number has resulted in sometimes positive, negative, or null relationships between species richness and ecosystem functioning (Grace et al., 2007; Roscher et al., 2012).

Lastly, because trait-based ecology measures properties of individuals that are linked to the environment and because it attempts to make predictions for ecosystem functioning, it must be able to scale from individuals to ecosystems. However, achieving this requires an exciting but extremely challenging synthesis of physiology, population biology, evolutionary biology, community ecology, ecosystem ecology, and global ecology (Reich, 2014; Webb et al., 2010). In this chapter, we suggest combining trait-based approaches with MST to make some progress on this problem.

Here, we present a novel theoretical framework to scale from traits to communities to ecosystems and to link measures of diversity. We argue that trait-based ecology can be made more predictive by synthesizing several key areas of research and to focus on the shape and dynamics of trait distributions. Our approach is to develop more of a predictive theory for how environmental changes, including land use and shifts in abiotic factors across geographic and temporal gradients, influence BEF (Naeem et al., 2009). We show how starting with a few simple but general assumptions allows us to build a foundation by which more detailed and complex aspects of ecology and evolution can be added. We show how our approach can reformulate and generalize the arguments of Chapin et al. (2000), McGill et al. (2006) and Violle et al. (2014) by integrating several insights from trait-based ecology (Garnier and Navas, 2012) and MST (Enquist et al., 1998; Gillooly et al., 2001; West et al., 1997). In doing so, we can derive a more synthetic theory that can begin to: (i) assess differing assumptions underlying the assembly of species; (ii) assess the relative importance of hypothesized drivers of trait composition and diversity; and (iii) build a more predictive and dynamical framework for scaling from traits to communities and ecosystems. We call this theory, Trait Driver Theory or TDT, because it links how the dynamics of biotic and abiotic environment then drive the performance of individuals and ecosystems via their traits. Combining MST with trait driver approaches allows TDT to work across scales and also addresses one of MST's key criticisms: it does not incorporate ecological variation—such as trait variation—and cannot be applied to understanding the forces that shape the diversity and dynamics of local communities (Coomes, 2006; Tilman et al., 2004).



2. TRAIT DRIVER THEORY

TDT is based on a synthesis of three influential bodies of work. The first are trait-based approaches that are largely encapsulated in Grime's Mass Ratio Hypothesis or MRH (Grime, 1998). The MRH states that ecosystem functioning is determined by the characteristics or traits of the dominant (largest biomass) species. Implicit in the MRH is the idea that traits of the dominant species are a more relevant measure than species richness. The second component is the generalized and quantitative approach to trait-based ecology through Norberg et al. (2001) who used a mathematical framework to link the distribution dynamics of phenotypic traits with environmental change and ecosystem functioning (Norberg, 2004; Norberg et al., 2001; Savage et al., 2007; Shipley, 2010). The third component is Metabolic

Scaling Theory or MST. MST can be used to predict how variation in organismic size and the traits associated with metabolism will then influence individual performance (growth and resource use), and how these performance measures will then scale up to influence populations, communities, and ecosystems (Enquist et al., 1998, 2003, 2009; Savage et al., 2004; Yvon-Durocher et al., 2012). MST achieves this by showing how variation in individual rates of mass growth, dM/dt , and metabolism, B , can be linked to variation in a few key traits (e.g. body size, M , and traits related to cellular metabolism and allocation; see Enquist et al., 2007b, 2009; von Allmen et al., 2012; West et al., 2002).

2.1 The Central Assumptions of Trait-Based Ecology and the ‘Holy Grail’ of Trait-Based Ecology

Trait-based ecology assumes that there are traits that are functional, meaning they link the environment to variation in whole-organism performance and ultimately fitness (Schmitz et al., 2015; Violle et al., 2007; see Fig. 1). That is, as shown in Fig. 1 and as described in Schmitz et al. (2015 in this special issue), variation in traits influences organismal performance (e.g. metabolism, growth rate, demographic rates) and ultimately fitness (Ackerly and Monson, 2003; Garnier et al., 2004; Lavorel et al., 2007; Violle et al.,

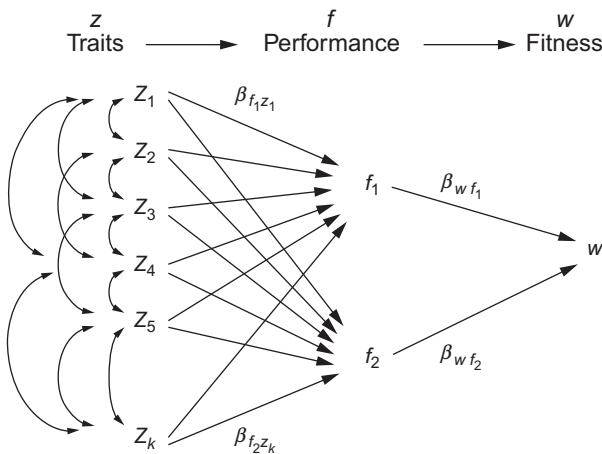


Figure 1 Path diagram representing the linkages between phenotypic traits, z , performance measures, f , and fitness, w . The arrows represent possible mechanistic linkages between traits. The coefficients β represent the correlation coefficients between traits, z , functions, f , and ultimately fitness, w . *Note:* performance measures include growth rate as well as survivorship and reproductive rates. Metabolic scaling theory explicitly links traits to these performance functions. Trait Driver Theory aims to link the performance function with community assembly and ecosystem functioning. *Figure modified from Kingsolver and Huey (2003) and Arnold (1983).*

2007). This approach has been recently validated with a comparative study linking variation in individual-level traits with variation in life history and demography parameters (Adler et al., 2014). Another key assumption of trait-based ecology is that traits of individuals can be used to predict individual performance that can be effectively summed or scaled up to the functioning of ecosystems (Lavorel and Garnier, 2002; Suding et al., 2008b). The *raison d'être* and the ‘Holy Grail’ of trait-based ecology is to use functional traits, rather than species identities, to better predict community and ecosystem dynamics (Lavorel and Garnier, 2002; Lavorel et al., 2007; Suding and Goldstein, 2008).

2.2 Linking Traits, Individual Performance, Communities, and Ecosystem Functioning

We start by extending Grime’s MRH. Grime argued that dominant traits rather than species number drive ecosystem functioning. As a result, it is crucial to measure the trait frequency distribution defined by biomass for the assemblage. An important question is in order to assess the MRH should one use abundance- or biomass-weighted mean trait values to best estimate the frequency distributions. In Section 5 below we further address this question, however, to start TDT also focuses on the trait frequency distribution $C(z)$ —the histogram of biomass across *individuals* characterized by a given trait value, z , summed across all individuals within and across species. Thus, $C(z)$ captures both intra- and interspecific trait differences. However, unlike the MRH and current emphasis on using community-weighted mean traits, we are interested in the overall shape of the distribution of phenotypes of individuals as measured by the central moments—variance, skewness, kurtosis—beyond the mean. We can link individual growth rate and the population *per capita* growth rates via how traits influence organismal performance via the growth function,

$$f(z) = \left[\frac{1}{C(z)} \right] \left[\frac{dC(z)}{dt} \right] \quad (1)$$

where $C(z)/dt$ is the biomass growth rate for *all* individuals with a given trait value z (see [Appendix](#)). By integrating the growth equation across all values of the trait across individuals, we can derive dynamic equations for how total community biomass, C_{Tot} , depends on the shape of the biomass-trait distribution, $C(z)$, and how that shape itself changes in time. Consequently, the net production of biomass in the community or Net Primary Productivity (NPP) is

$$\frac{dC_{\text{Tot}}}{dt} = \int f(z)C(z)dz \quad (2)$$

(see also [Lavorel and Garnier, 2002](#); [Norberg et al., 2001](#); [Vile et al., 2006](#)). Equation (2) requires understanding what sets the form of $f(z)$.

2.3 Linking Dynamics of Trait Distributions to Environmental Change and Immigration

Starting with [Norberg et al. \(2001\)](#) and [Savage et al. \(2007\)](#), we focus on how traits that strongly influence organismal growth rate are influenced by the environment, E . In [Fig. 2](#), we show an example of how variation

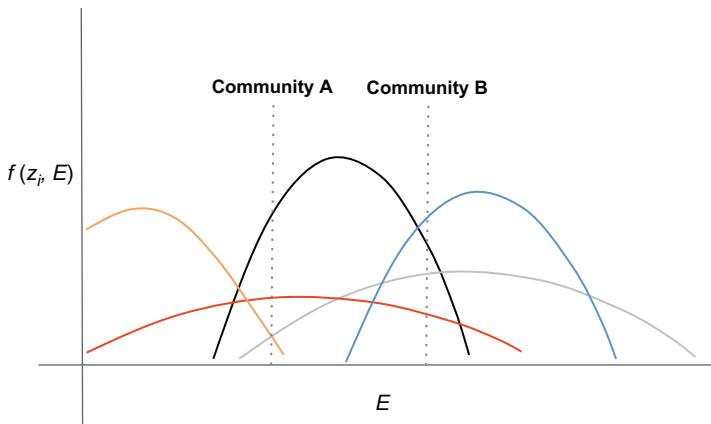


Figure 2 Graphical representation of essential components of Trait Driver Theory. Each curve represents positive performance (here shown as the growth function f) or growth rates of individuals characterized by a unique trait value, z . Each colour then indicates how a given trait value then translates to variation in organismal growth rate across an environmental gradient, E . Because of a trade-off between traits and the environment (see Section 2.3), each trait has an optimal environment where growth is maximized, and each trait exhibits a unimodal response to an environmental gradient, E . At different points along an environmental gradient, different traits are characterized by the highest growth rate, f . Also, species can differ in the width of their performance curves, σ_z^2 . Here in two communities, A and B, although several trait values can achieve positive growth, in community A the black phenotype has the highest growth rate in that location and is predicted to then be the most dominant phenotype in a with trait z . In contrast, in community B, the black phenotype is predicted to not be as dominant. This trait value then is the optimal trait value, given the potential species pool, for that community. *Note:* in this example, growth rates are only influenced by E . However, extensions of TDT (see text) can assess how the growth rate of a given trait value then is influenced by the presence and dominance of other trait values. Such trait–trait interactions could then modify the shape and breadth of each growth curve.

in a given trait, z , translates to variation in *per capita* growth rate across an environmental gradient. This example assumes that all individuals in a given community ultimately compete for similar limiting resources, and that there is an optimal environment where growth is fastest (Fig. 2). Although our approach starts with a single trait, trait-based models can straightforwardly incorporate multiple, correlated traits (Savage et al., 2007; see also Appendix). By incorporating temporal environmental forcing into the growth function, TDT predicts how the distribution of traits, $C(z)$, responds to both biotic and abiotic drivers (Norberg et al., 2001; Savage et al., 2007).

The shape and the dynamics of the trait distribution ultimately reflect a balance between two rates—the introduction/immigration of traits, I , into an assemblage and the outcome of variation in the performance, f , of those traits within the assemblage. A general trait-based equation for growth and immigration is given by

$$\frac{dC(z)}{dt} = f[z, E, C(z)]C(z) + I[z, E, C(z), C_{\text{landscape}}(z)] \quad (3)$$

Here, we now explicitly include the effects of the environment, E , and the trait distribution $C(z)$ as part of the growth, f , and immigration, I , functions because they can influence both via environmental change, competition, facilitation, sampling effects, or other biological interactions such as density dependence (Savage et al., 2007). The second term, the immigration function, I , reflects the external input of individuals into the community stemming from dispersal as well as the introduction of traits into the assemblage from other factors including evolutionary processes (mutations) and potential seed banks. The immigration function, I , reflects dispersal behaviour of individuals and is a function of other ecological factors, such as organismal traits, and may be influenced by the abundance of that trait already in the local community $C(z)$ as well as the abundance of that trait in the larger landscape, $C_{\text{landscape}}(z)$ (see Gilbert and DeLong, 2015; Laskowski et al., 2015; Schmitz et al., 2015 in this issue).

Next, we use two assumptions to constrain the form of Eqs. (2)–(3). First, a central tenet of TDT and a well-grounded concept in ecology and evolution are that across an environmental gradient, E , organisms will tend to have a unimodal functional response in their performance and fitness functions (Fig. 2). As a result, a shift in the environment, E , will affect the *per capita* population growth rate and thus the traits that are dominant in the community or assemblage (Davis and Shaw, 2001; Whittaker et al., 1973) and the rate of trait evolution (Levins, 1968). Second, there

are specific traits that link environmental drivers to individual growth rate, and the trait driven *per capita* biomass growth rate, $dC(z)/dt$ (Fig. 1; Arnold, 1983). The performance or growth function $f(z)$ then is a result from an environment-mediated trade-off between traits, such as investment in growth versus defence or from investment in growth rate versus desiccation resistance. As a result, for a constant environment, E , there are optimal trait values, z_{opt} , that maximize the growth function given in the environment.

In the case of a single-trait optimum, we approximate this as a symmetric function such as a Gaussian or quadratic trade-off $f(z) \propto \left[1 - \left(\frac{z - z_{\text{opt}}}{\sigma}\right)^2\right]$, where σ^2 is the trait breadth of the trade-off function. If the environment is constant and immigration, I , is zero, individuals with traits that match, z_{opt} , will gradually replace all other individuals, and the trait distribution will collapse on a single point for the optimal trait value z_{opt} (Norberg et al., 2001). Thus, TDT is consistent with a competitive trait hierarchy view of assemblage interactions (Freckleton and Watkinson, 2001; Goldberg and Landa, 1991; Kunstler et al., 2012; Mayfield and Levine, 2010) as well as a population source-sink view of assemblage (Pulliam, 1988) and metapopulation perspective of trait dynamics across environmental gradients (Davis and Shaw, 2001). As we discuss below, additional biotic and abiotic interactions and processes can also be shown to influence the shape of the trait distribution via the growth and immigration functions (see Weiher and Keddy, 1995).



3. PREDICTIONS OF TDT

Next, we emphasize the central predictions of TDT. These predictions are also summarized in Table 1 in terms of how different measures of the trait distribution can provide novel insight and predictions regarding the main drivers of the current composition of the species assemblage as well as the future dynamics of the species assemblage.

Prediction (1): Shifts in the environment will cause shifts in the trait distribution (Fig. 3; Table 1).

Prediction (2): The difference between the optimal trait and the observed mean trait, as well as the trait variance, provides a measure of the capacity of a community to respond to environmental change (Figs. 3 and A1; Table 1).

Shifts in the abiotic or biotic environments, represented by E and $C(z)$, respectively, will lead to corresponding shifts in the community trait distribution. The magnitude of the shift over some time and the rate of change of

Table 1 Summary of the Core Predictions from Trait Driver Theory for How the Different Central Moments of the Trait Distribution Will Respond to Differing Biotic and Abiotic Forces and How They Will Then in Turn Influence Community Dynamics and Ecosystem Functioning

Moment of Community Trait Distribution, $C(z)$	Predictions for Rate of Community Response to a Changing Environment	Predicted Ecosystem Effects
I. Mean	<p>(a) Will shift if environmental change alters value of z_{opt} and time scales are not too rapid and oscillatory</p> <p>(b) Lags z_{opt} by an amount that depends on rate of change in environment, rates of immigration, and the forces that influence the variance</p>	<p>(i) Will shift productivity according to form of growth equation, f</p>
II. Variance	<p>(a) Decreases with strong abiotic filtering</p> <p>(b) Decreases due to competitive exclusion by individuals with trait z_{opt}</p> <p>(c) Can increase with increased immigration, competitive niche displacement, and/or temporal variation in z_{opt} due to a variable environment</p> <p>(d) Under neutral theory, if no immigration or mutation, variance will decrease over time so as to decrease response abilities over time</p>	<p>(i) Increased variance implies lower productivity for fixed or stable environment</p> <p>(ii) Increased variance accelerates community response to environmental changes</p> <p>(iii) Increased variance will lead to increased stability of ecosystem functioning by reducing the lag of \bar{z} and z_{opt} in varying environments</p>
III. Skewness	<p>(a) Skewness values $>$ or $<$ 0 can reflect a lag between \bar{z} and z_{opt} and a rapidly changing community due to an environmental driver or extreme limit to a trait value</p> <p>(b) Increases in skewness can indicate a response to rapid environmental changes or the importance of rare species advantages in local coexistence</p>	<p>(i) Depending upon kurtosis and variance value, productivity should be reduced compared with a community with similar variance but skewness equal to zero</p>

Continued

Table 1 Summary of the Core Predictions from Trait Driver Theory for How the Different Central Moments of the Trait Distribution Will Respond to Differing Biotic and Abiotic Forces and How They Will Then in Turn Influence Community Dynamics and Ecosystem Functioning—cont'd

Moment of Community Trait Distribution, $C(z)$	Predictions for Rate of Community Response to a Changing Environment	Predicted Ecosystem Effects
IV. Kurtosis	<p>(a) Positive kurtosis – a more peaked distribution – reflects competitive exclusion or other types of biotic exclusion</p> <p>(b) Kurtosis close to -1.2 reflects a uniform distribution consistent with uniform niche partitioning</p> <p>(c) More negative values could reflect the coexistence of contrasting ecological strategies, recent or sudden environmental change</p>	<p>(i) If the trait mean equals z_{opt}, forces that decrease kurtosis will decrease productivity while forces that increase kurtosis will increase productivity</p> <p>(ii) In a temporally varying environment, more negative kurtosis values will lead to increased stability of ecosystem functioning by reducing the lag of \bar{z} and z_{opt}</p>

Trait Driver Theory (TDT) can incorporate each of these forces via the shape of the trait biomass distribution, $C(z)$, to then make specific predictions for how each can drive the dynamics of $C(z)$ and ecosystem functioning (see text). Parameterizing predictions for specific cases depends upon the traits that affect the growth rate, f . Here \bar{z} is the average observed trait value and z_{opt} is the optimal trait value for an given environment, E .

the shift can both be calculated from Eq. (2). According to Eq. (2), the value of the optimal trait will change with the environment (e.g. Ackerly, 2003), while the mean trait of the community will approach the optima but with a lag in time according to how long it takes for either trait plasticity and/or the processes of species sorting and selection to act (Ghalambor et al., 2007). In environments where the optimal trait value is changing quickly relative to generation times or plasticity, there may be little capacity for the mean community trait, \bar{z} , to track these changes. In such circumstances, \bar{z} , may never, or only rarely, be expressed at an optimal value for the current environment. Nonetheless, we expect that for most communities, the difference between the optimal trait, $z_{\text{opt}}(E)$, and the observed mean trait, \bar{z} , or $\Delta(E)$, will be a measure of how the community has responded/will respond to environmental change. Norberg et al. (2001) derive the general expression

$$\Delta(E) = z_{\text{opt}}(E) - \bar{z} \quad (4)$$

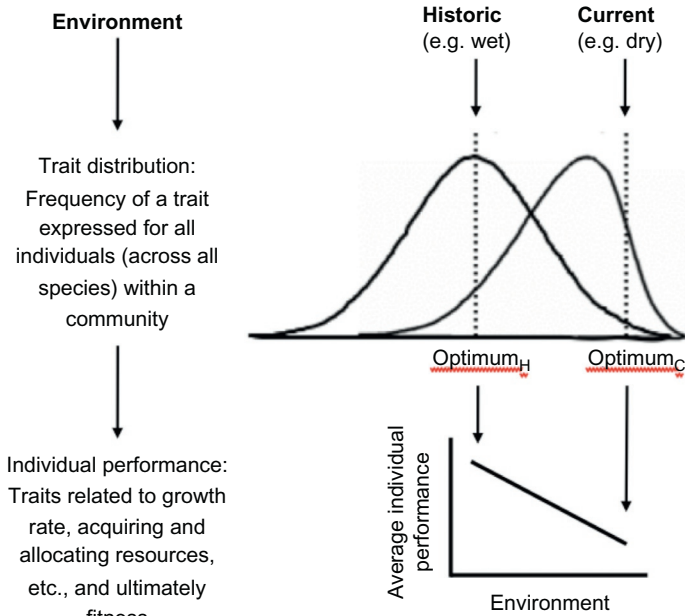


Figure 3 Conceptual diagram linking changes in optimal traits in changing environments with the frequency distributions for that trait in historic versus current environments. In this example, the optimal trait value (dotted line) has shifted to the right. Individual performance (growth, fitness, etc.) is highest at the optimal trait expression in an environment. *Note:* because the response of an assemblage cannot be instantaneous, the trait distribution has an increased skew. Further, since the highest trait frequencies are not yet at the optimal trait expression, the average performance in the current environment is lower than in the historic environment.

where $\Delta(E)$ quantifies the community trait ‘lag’ in relation to the current environment. This measure is analogous to the ‘lag load’ in evolutionary theory (Maynard Smith, 1976). We can thus define $d\Delta/dt$ as the response capacity of a community. Equation (4) predicts that capability of the assemblage to respond to directional shifts in the environment will be directly proportional to the trait variance, $d\Delta/dt \propto V$. Importantly, within TDT, directional selection for a given optimal trait value need not always lead to an increase in *per capita* growth rate, f . Because of trade-offs between traits and frequency and density-dependent effects on performance and fitness, the performance (and fitness) associated with the new optimum value likely differs from the fitness and growth rate in the previous environment (Antonovics, 1976; Dieckmann and Ferrière, 2004; Ferrière and Legendre, 2013). For example in Fig. 3, we highlight a hypothetical example of a shift in the community trait distribution from wet to dry that comes with

a decrease in optimal performance. Extensions of TDT can in principle include these effects (Savage et al., 2007).

In sum, predictions 1 and 2 formalize Chapin et al.'s conceptual framework (Chapin et al., 2000) for the development of a predictive trait-based ecology. In the case of multiple traits underlying growth, f , differing trait combinations could lead to similar growth rates in differing environments (see also Marks and Lechowicz, 2006). We note that these predictions implicitly ignore the effects of frequency dependence but elaborations of TDT can include these effects (see equation 5 in Savage et al., 2007).

Prediction (3): The skewness of the trait distribution can be an indicator of past or ongoing immigration and/or environmental change due to lags between growth, reproduction, and mortality (Table 1).

Because of time lags between environmental change and the time scale of organismal responses (growth, demography, etc.), the trait distribution of an assemblage will not be able to instantaneously track environmental change, and skewness in the trait distribution will develop (see also Figs. 3 and A1). Alternatively, skewness may reflect differential immigration of traits from one side of a habitat or a community that contains 'sink' populations (Pulliam, 1988) supported via immigration (see also Pawar, 2015 in this issue). Neutral theory (Hubbell, 2001), in which traits have no demographic effects, could also lead to skewed distributions due to neutral trait evolution. The implication is that trait-based ecology can infer the dynamics of trait assemblages via assessing the shape of contemporary trait distributions. Combining information on the shape trait distributions with additional information such as dispersal history and/or size distributions would help separate lag effects from drift and differential immigration.

Prediction (4): The rate of change of net ecosystem productivity in response to environmental change can be predicted via the growth function, f , and the shape of the community biomass-trait distribution $C(z)$ at some initial time (Table 1).

In the simple case of a single trait with a single environmental driver, Norberg et al. (2001) derived a general expression linking the dynamics of the trait distribution by noting Eq. (2), Eq. (3) can be approximated as

$$\frac{dC_{\text{Tot}}}{dt} \approx \left[f(z, E, C(z))_{z=\bar{z}} + \frac{\partial^2 f(z, E, C(z))}{\partial z^2} \Big|_{z=\bar{z}} V \right] C_{\text{Tot}} + I \quad (5)$$

Equation (5) follows from a Taylor expansion that effectively linearizes the equations. If the terms in brackets depend on total biomass, dC_{Tot}/dt would scale non-linearly with total biomass, but in the simplest case, these

terms are independent of total biomass implying that production scales linearly with total biomass. In Eq. (5), the net primary production, C_{Tot}/dt , is equal to the growth rate of the mean community trait, \bar{z} , plus the second term that accounts for how much variation there is in the community trait distribution, V . Because the growth function, $f(z, E)$, has a maximum at z_{opt} , we expect the second derivative term to be negative, as long as the average observed trait value, \bar{z} is in the neighborhood of the optimal trait value, z_{opt} (Norberg et al., 2001; see also discussion in Appendix) reflecting the increasing reduction in growth rate as trait values increasingly differ from z_{opt} (see Fig. 2 and Eq. 4). The unimodal shape is the simplest assumption requiring only the mean and variance. There is reason to expect that f can be approximated as unimodal. For example, growth rates typically exhibit unimodal response with measures of temperature, pH, etc. (McGill et al., 2006). Again, the term I gives the addition of biomass through immigration/dispersal.

Prediction (5): Within a community whose growth rate depends on a single trait, an increase in the variance of that trait will lead to a decrease in net primary production (Table 1).

An additional prediction is that for communities whose growth is driven by a single key trait, larger trait variance, V , will decrease the net primary production for the whole community because a higher proportion of individuals differ from z_{opt} (Norberg et al., 2001). This idea of a trade-off between short-term productivity and long-term response to environment is reflected by agricultural imperatives with agricultural issues, where short-term productivity is emphasized and variance in traits is minimized in trait values and short-term productivity is maximized. Elaborations of TDT have shown that incorporating multiple limiting resources, multiple traits, and trait covariation (Savage et al., 2007) can weaken, nullify, or even reverse the predicted negative relationship between dC_{Tot}/dt and V .

From Eqs. (3) and (4), the rate at which a community can track environmental change will be greater when there is greater trait variance. Intuitively, greater variance leads to more extreme traits being immediately available to respond to environmental change. Thus, the rate of response will also depend upon the specific form of the growth function f (e.g. for a given value of z , how does f vary across an environmental gradient? see Savage et al., 2007 and discussion in Appendix). For example, if f is a simple Gaussian or polynomial function with E (Fig. 1), then the value of $d\Delta/dt$ can be approximately proportional to the community trait variance, V . Building on the work of Norberg et al. (2001) and Savage et al. (2007), these equations can be extended to include higher-order moments such as skewness and kurtosis.

3.1 Extending TDT via Recasting and Assessing Different Ecological Hypotheses About Diversity and Trophic Interactions

TDT predicts that over time only one phenotype should dominate a given environment. An important question is what maintains diversity (trait variation) within an assemblage? According to TDT trait, variance can be increased by many different ways (also see [Dell et al., 2015](#); [Gilbert and DeLong, 2015](#); [Pawar, 2015](#) in this issue; [Pettorelli et al., 2015](#)). Immigration, I , from outside the assemblage, as well as from a directionally shifting or a temporally variable environment ([Norberg et al., 2001](#); [Savage et al., 2007](#)), can increase and/or maintain trait variation. Further, theoretical elaborations of TDT have shown that the diversity of phenotypes (traits) present in a given assemblage can be influenced by trade-offs between traits that influence growth. For example, trade-offs between allocation to predator defence and growth rate ([Norberg et al., 2001](#); [Savage et al., 2007](#)) can increase the variance of a given trait. In a variable environment, correlations between traits that underlie the growth function, f , lead to the survival of organisms with trait values that are less favourable in the current environment but may be well suited for new environments that arise. Thus, phenotypic trait correlations among traits can ramify to have quantitative effects on ecosystem dynamics (lowering NPP) and enable assemblages to better track environmental change ([Savage et al., 2007](#)).

Additionally, trait variation can also stem from additional ecological hypotheses for biological diversity. An exciting aspect of TDT is that differing ecological hypotheses based on species richness can be recast in terms of traits. In [Table 1](#) and [Fig. A1](#), we overview the predictions of the different theories as recast in the light of TDT. As a result, TDT can then be used to ‘scale up’ the implications of many differing classic and current hypotheses for species richness via the assumptions of trait distributions implicit in these theories. Another exciting potential of TDT is to show (i) how different trophic interactions and variation in traits controlling ‘predation risk’ and ‘prey selection’ would then influence community trait and size distributions (see [Pettorelli et al., 2015](#) in this issue) and (ii) how individual trait variation could potentially have opposing effects on predator–prey dynamics (see [Gilbert and DeLong, 2015](#) in this issue). TDT would then offer a framework to show how these differing ecological processes then scale up to influence ecosystem functioning.

As we show in Appendix A.1 we can use TDT to recast several different ecological theories in terms of the distribution of functional traits. Different

ecological theories based on species richness (neutral theory, abiotic filtering, competitive exclusion, Chesson's storage effect, or rare species advantages; see discussion in [Appendix](#)) as well as the effects of abiotic processes (shifts due to environmental change, disturbance) will uniquely influence the shape of the community trait distribution and the potential of the assemblage to maintain diversity, as reflected in changes in the trait variance, dV/dt . The relative strengths of abiotic, biotic, and neutral processes will lead to different shapes of trait distributions that will have different implications for responses of community to directional shifts in the environment as well as ecosystem functioning.



4. EXTENDING AND PARAMETERIZING TDT

4.1 Scaling from Individuals to Ecosystems Using MST

So far, TDT assumes that there is no variation in organismal size. Instead, the total biomass associated with a trait, z , is denoted by $C(z)$. This notation avoids ever needing to account for *individual organismal* mass, M , or even the number of individuals with mass. However, body size can vary greatly—it is also an important trait that influences variation in organismal metabolism ([Peters, 1983](#)), population growth rate ([Savage et al., 2004](#)), and abundance ([Damuth, 1981](#); [Enquist et al., 1998](#)). The scaling equations in MST differ from TDT so far as they are phrased in terms of individual mass. In order to integrate these theories, we use three insights from MST ([Enquist et al., 2007b, 2009](#); [Savage et al., 2004](#); [West et al., 1997, 2009](#)) to explicitly formulate TDT to work across scales in organismal size, M , and environmental changes or gradients in temperature, T . As we show, MST provides the basis to formally link traits, organismal growth rate, and ecosystem fluxes ([Enquist et al., 2007a,b](#)).

First, MST is explicit about how the organismal growth is dependent upon the size of the organism. In the case of MST, we start with how organismal biomass growth rate, dM/dt , is related to whole-organism metabolic rate, B , and organismal mass, M , as

$$\frac{dM}{dt} = b_0(z)M^\theta \quad (6)$$

where $b_0(z)$ is a metabolic coefficient that depends on a single or set (meaning z is a vector) of traits. The allometric scaling exponent θ is hypothesized to reflect the branching geometry of vascular networks ([Enquist et al., 2007b](#)). Theory and empirical data point to $\theta \approx 3/4$ for large size ranges ([Enquist et al., 2007c](#); [Savage et al., 2008](#)). Equation (6) has recently been

shown to be a good characterization of tree growth (Stephenson et al., 2014) and is a specific case of a more generic growth function (Moses et al., 2008; West et al., 2001) that can be applied to both plants and animals. While we focus here on a specific plant growth model, we note that other trait-based models have recently been developed for animals and phytoplankton (Litchman and Klausmeier, 2008; Muller et al., 2001; Ricker, 1979; West et al., 2001), and they could also be used to parameterize TDT. Below, we elaborate Eq. (6) to explicitly include the traits for plants that underlie b_0 and how we can use this equation as the basis for a general trait-based growth function.

To integrate MST into TDT, we first recognize that, $C(z)$, the biomass associated with trait z can be expressed as $C(z) = \int dM C(z, M) = \int dM N(z, M)M$, where $C(z, M)$ is the mass density of individuals with both trait value z and individual mass M , while $N(z, M)$ is the number density of individuals that have both trait value z and individual mass M . In this expression, we have integrated over all possible values of mass, M , so that we have the total biomass of *all* individuals with trait z . Furthermore, note that integrating this over all traits, z , gives the total biomass, $C_{\text{Tot}} = \int dz C(z) = \int dz \int dM N(z, M)M$.

To integrate MST into TDT, we solve for the conditions of steady state where $N(z, M)$ is not changing in time. It can be shown (see Appendix) that the equation for the scaling of NPP with the total biomass of the assemblage is,

$$\frac{dC_{\text{Tot}}}{dt} = \left\langle b_0(z)M^{-1/4} \right\rangle_C C_{\text{Tot}} \quad (7)$$

where dC_{Tot}/dt scales isometrically with C_{Tot} and the C subscript denotes that the average, denoted by brackets $\langle \cdot \rangle$, is taken with respect to the biomass. This equation is in the most generic form of a general TDT equation. Note that the TDT growth function, f , is now

$$f(z) = b_0(z)M^{-1/4} \quad (8)$$

and can be expanded and expressed in terms of the biomass-weighted central moments of the trait z , such as the variance, skewness, and kurtosis (see below). Again, the exponent, $-1/4$ is the idealized case and empirical values that may deviate from $\theta \approx 3/4$ can be used.

4.1.1 Incorporating Environmental Trade-Offs in the Growth Function, f

Equation (8) alone would predict that the *per capita* growth rate will increase forever as the trait, b_0 , and mass-specific metabolic rate increase. In reality,

though, there is some range of trait values at which the organism can grow. This is because there are trade-offs in performance and fitness: decrease in growth when, for a given value of E , the trait value gets either too small or too large. In the case of a given leaf trait such as leaf size or leaf investment (closely associated with variation in photosynthetic rates and the specific leaf area (SLA)), at some point, continued increases in leaf nitrogen may ultimately limit resource uptake as high N would result in individuals more prone to herbivores, pathogens, etc. and/or will result in water transport demands that would increasingly be maladaptive for a given local environment. Ultimately, one cannot have an infinitely large leaf, an infinitely thin leaf, or a plant that is all leaf area. Thus, deviation away from $b_{0,\text{opt}}$ would be associated with a trade-off between specific trait values and plant performance (such as growth rate, survivorship, and/or reproduction; see also [Ghalambor et al., 2007](#)).

Incorporating trade-offs between trait values, the environment, and performance is central to TDT. We can incorporate these trade-offs in a general form by multiplying the scaling relationship by a quadratic function. As a result, $f(b_0)$ is maximal at the optimal trait, $b_{0,\text{opt}}$, and the niche width defined by $\sigma_{b_0}^2$ where

$$f(z) = b_0(z)M^{-1/4} \left(1 - \frac{(b_0 - b_{0,\text{opt}})^2}{\sigma_{b_0}^2} \right) \quad (9)$$

Here, the second term is the trade-off function, and c_0 is an overall constant coefficient. Expressing $f(b_0)$ across an environmental gradient, $f(b_0, E)$, would then reveal a unimodal growth function ([Fig. 2](#)).

The second insight from MST shows that the metabolic normalization, b_0 , can be linked to specific traits. For the plant growth function, building on the insights from the relative growth rate literature ([Evans, 1972](#); [Lambers et al., 1989](#); [Poorter, 1989](#)), [Enquist et al. \(2007b\)](#) derived an extension to Eq. (6) that explicitly details the traits that together define b_0 and hence f so that

$$b_0 \propto \frac{c}{\omega} \left(\frac{a_L}{m_L} \right) \dot{A}_L \beta_L \quad (10)$$

Equation (11) shows that, in addition to plant size, M , the rate of growth is governed by the scaling exponent, θ , and five traits: (i) \dot{A}_L , the net leaf photosynthetic rate (grams of carbon per area per unit time); (ii) a_L/m_L , the SLA, the quotient of area of the leaf, a_L , and the mass of a leaf, m_L ;

(iii) ω , the carbon fraction of plant tissue; (iv) ϵ , the carbon use efficiency of whole-plant metabolism, and (v) β_L , the leaf mass fraction (the ratio of total leaf mass to total plant mass) which is a measure of allocation to leaves. As a result, we can parameterize TDT with specific traits that underlie b_0 .



5. ADDITIONAL PREDICTIONS OF TDT

In the second column of [Table 1](#), we summarize additional TDT predictions for scaling up community or assemblage trait distributions to predict several ecosystem-level effects. Specifically, the shape of the trait distribution as measured via the central moments of the distribution.

Prediction (6): Ecosystem net primary productivity, dC_{Tot}/dt , will scale with the total biomass but will be influenced differently by the mean and variance of the community trait distribution.

A third insight from MST allows us to more formally link TDT with MST by including organismal mass dependence into TDT. In particular, most assemblages of organisms will be characterized by a distribution of sizes. For plants, following the arguments in [Enquist et al. \(2009\)](#), we can substitute the distribution of the number of individuals as a function of their size, M , or the size spectra, $N(M)$. For the idealized case of $\theta \approx 3/4$, they show that $N(M) \propto M^{-11/8}$ and link the total biomass, C_{Tot} , with the size of the largest individual, M_b , where $M_b \propto C_{\text{Tot}}^{8/5}$. This allows us to consider a few special cases of [Eq. \(7\)](#) that relate TDT and scaling equations already in the literature.

In the case of a given assemblage where there is no size distribution and only a single mass value, M^* , or a very small range of mass values, the scaling of NPP becomes

$$\text{NPP} = \frac{dC_{\text{Tot}}}{dt} = (M^*)^{-1/4} \langle b_0(z) \rangle_C C_{\text{Tot}} \quad (11)$$

The term $(M^*)^{-1/4}$ can be thought of as an overall normalization to the growth function $f(z)$ from TDT. As such, this result reveals that TDT, as originally formulated (see [Eq. 3](#)), ignores variation in individual mass. Thus, based on [Eq. \(6\)](#), growth functions within TDT should have a roughly $(M^*)^{-1/4}$ hidden with the normalization constant for their growth function. In the case where (i) organisms within the community or assemblage can differ greatly in their sizes; (ii) z and M are uncorrelated; and (iii) the number density is a separable function, such that $N(z, M) = N(M)N(z)$; it can be

shown that the growth equation can be expressed in two different ways. Each way depends on how one averages the trait distribution. In the first case, we have

$$\frac{dC_{\text{Tot}}}{dt} = k \langle b_0(z) \rangle C_{\text{Tot}}^{3/5} \quad (12)$$

and in the second case, we have

$$\frac{dC_{\text{Tot}}}{dt} = k \langle b_0(z) \rangle \langle M^{-1/4} \rangle_C C_{\text{Tot}} \quad (13)$$

where k is a proportionality constant. Equation (13) is equivalent to Eq. (12) but expresses the growth function more in terms of the TDT framework such that the right side appears to have an overall *linear* dependence in C_{Tot} . As a result, in Eq. (12) we have a mixture of types of averages, with $\langle b_0(z) \rangle$ being the abundance average of the function $b_0(z)$, while $\langle M^{-1/4} \rangle_C$ is the biomass average of $M^{-1/4}$. Both equations are equivalent ways to express the scaling of NPP function. Equation (12) is a more simple expression and only involves using the abundance average of the trait distribution. Equation (12) consolidates the organism mass average with the 3/5 scaling dependence of C_{Tot} . These derivations help to clarify when trait-based studies should use biomass- or abundance-weighted values.

Both equations assume a community steady-state approximation where $N(z, M)$ is not changing in time. If this is violated (e.g. the community trait abundance or number distribution $N(z, M)$ is changing), then deviations from Eqs. (12) and (13) are expected. Nonetheless, these equations provide a basis for linking the scaling of organismal growth rate and trait variation of individuals with ecosystem-level processes. For all of these equations and cases, the functions inside the averages can be expanded in terms of moments as done for TDT for biomass-weighted averages or as done in Savage (2004) for abundance-weighted averages.

Putting all of this together with Eq. (5) yields the prediction that Eqs. (12) and (13) are then modified by the shape of the trait distribution, where for a given E , growth is reduced with departure from $b_{0,\text{opt}}$,

$$\frac{dC_{\text{Tot}}}{dt} \approx k \langle b_0(z) \rangle + \left[\left(1 - \frac{(b_0 - b_{0,\text{opt}})^2}{\sigma_{b_0}^2} \right) \frac{d^2 f}{dz^2} \Big|_{z=\bar{z}} V(b_0(z)) \right] C_{\text{Tot}}^{3/5} \quad (14)$$

The second term captures the reduction of production due to deviation from b_{opt} . Equation (14) represents a formal integration of foundations of TDT from Norberg et al. (2001) with MST from West et al. (1997) and Enquist et al. (2009). Because the growth function, $f(z, E)$, has a maximum at z_{opt} (Fig. 2), we expect the second derivative term, d^2f/dz^2 , to be negative, as long as \bar{z} is in the neighbourhood of z_{opt} . Importantly, Eq. (14) enables one to parameterize TDT with a specific trait-based growth function. Further, it enables the integration of physiological performance curves for how the key integrative trait, b_0 , varies across a given environmental gradient, E . Equation (8) predicts that dC_{Tot}/dt will increase with increasing community biomass, C_{Tot} . Note, here, the role of the relative breadth of species performance curves (see Fig. 1) is represented by $\sigma_{b_0}^2$.

Prediction (7): Equation (14) generates specific and testable relationships for the scaling of trait means, dispersion, and ecosystem production (see Table 3).

Equation (14) predicts that there is a range of mean trait values for which increases in the variance of traits, $V(b_0)$, will *decrease* Net Primary Productivity, dC_{Tot}/dt , and shifts in $\langle b_0 \rangle$ will lead to corresponding shifts in dC_{Tot}/dt . This will occur whenever the mean trait value is near the maximum of the growth function, which should occur frequently because evolution is driving the mean trait to match the optimal trait with some lag time. However, there are also mean trait values for which increases in the variance of traits, $V(b_0)$, will *increase* Net Primary Productivity (NPP), dC_{Tot}/dt , and shifts in $\langle b_0 \rangle$ will lead to corresponding shifts in dC_{Tot}/dt . This will happen when the mean trait value is further from the optimal trait value and below an inflection point in the growth function that occurs for small trait values (see the example of a shift from historically wet climate regime to a dry regime in Fig. 3). Intriguingly, this scenario suggests that trait variance can potentially act to either increase or decrease NPP depending on if the current trait distribution is close to the local trait optimum or not. So, if the assemblage is close to the optimal value, increases in trait variance will typically decrease NPP. This contrasts with biodiversity theories in which increasing variance (increased trait diversity) tends to increase NPP. Importantly, Eq. 14 shows the influence of variation, V , of the traits that underlie b_0 observed within the community.

Integrating MST into a more generalized TDT lists several key traits for TDT. We explore predictions of TDT in the special case of a single-trait driver such as SLA = (a_L/m_L) . First, a change in the environment will likely be associated with a shift in the mean value of SLA. Second, using Eqs. (12) and (14), we expect that

$$\frac{dC_{\text{Tot}}}{dt} \propto (\langle \text{SLA} \rangle - \langle V_{\text{SLA}} \rangle) C_{\text{Tot}}^{3/5} \quad (15)$$

Thus, according to Eq. (15), a shift that increases the abundance-weighted mean trait value of $\langle \text{SLA} \rangle$ will lead¹ to an increase in NPP. Preliminary support of this prediction comes from empirical studies that have noted that increases in the mean community SLA are closely linked with increases in ecosystem productivity (Garnier et al., 2004; Violle et al., 2007) and follows from TDT. Third, due to the productivity–variance trade-off predicted by TDT, holding the other variables constant, an increase in the community variance in SLA or V_{SLA} will lead to a *decrease* in productivity so long as the mean community SLA is close to the optimum value.

As we discuss below, TDT provides a foundation that can be modified by additional factors. When there are multiple trait shifts that may covary, measuring all of the traits listed in Eq. (11) would allow more detailed predictions. Of all the traits specified by Eq. (11), there appears to be evidence that SLA may vary more across environmental gradients than other traits and be more important for linking changes in a trait driver or environment, E , with variation in local plant growth (see Appendix). It varies across taxa (up to 3 orders of magnitude or approximately 1000-fold) and directionally varies across environmental gradients in soil moisture, irradiance, and temperature (see Garnier et al., 2004; Poorter et al., 2009; Wright et al., 2005). SLA has also been noted to vary considerably *within* species in response to local changes in climate and abiotic conditions (Cornwell and Ackerly, 2009; Jung et al., 2010; Shipley, 2000; Sides et al., 2014).



6. METHODS: ASSESSING TDT ASSUMPTIONS AND PREDICTIONS

6.1 Quantifying the Shape of Trait Distributions

In order to assess predictions of TDT, it is necessary to quantify the biomass distribution of traits, $C(z)$, in a species assemblage. This involves enough measurements of the trait values and body masses to obtain accurate

¹ Note: Because the growth equation f is for more instantaneous measures of growth, this prediction is based on rates of more instantaneous NPP and not necessarily annual net primary production. So, accurate testing of this prediction with annual productivity data should make sure to standardize for growing season lengths.

estimates of the underlying distributions, as guided by sampling theory and statistics (Baraloto et al., 2010; Paine et al., 2011). The sampling must occur across all individuals within our group and thus incorporates both inter- and intraspecific trait variability (see Appendix and Violle et al., 2012). The sampling protocols often make choices that limit accurate measurements of within-species variability more than across-species variability. Indeed, simultaneous measurements of intra- and interspecific trait measures are rarely collected (Ackerly, 2003; Baraloto et al., 2010). However, intraspecific variation in traits is important to determine the breadth of the distribution (Sides et al., 2014; Violle and Jiang, 2009). Trait abundance or biomass distributions, $C(z)$, can be approximated through sampling (see Appendix), so that predictions of TDT can be tested without explicitly measuring the traits of all individuals.

There are two reasonable approximations for community trait distributions. The first approximation method calculates the weighted trait distribution by taking the mean species trait value and multiplying by a measure of dominance (cover, biomass, abundance; Grime, 1998). This method can be implemented by calculating the central moments of the joint distribution. In the Appendix, we show the equations used to approximate the community trait moments in particular the community-weighted mean, variance, skewness, and kurtosis (CWM, CWV, CWS, and CWK). Community-weighted metrics, however, ignore intraspecific variation. Increasingly, it is becoming clear that intraspecific variation can contribute to a considerable amount of trait variation (Messier et al., 2010) and that relying on species mean trait values may not provide a robust measure of the shape of a trait distributions (Violle et al., 2012).

A second method utilizes sampling theories to help avoid the time-consuming work of sampling the traits of all individuals. Subsampling individuals can be used to better approximate how intraspecific variation influences the community distribution. In the Appendix, we develop this new method (see discussion in Appendix). The method utilizes subsampling individuals to obtain a better approximation of how intraspecific variation influences the community distribution. By subsampling individuals to estimate intraspecific trait variation within each species, one can begin to incorporate intraspecific variation around mean trait values for each species. We expect that utilizing this method in addition to the incorporation of how MST influences the scaling of the number of individuals will improve estimates for the shape of trait distributions. In short, by subsampling individuals for each species within a given assemblage, one

can begin to incorporate intraspecific variation around mean trait values for each species.

6.2 Testing Predictions of TDT

We tested several of the specific TDT predictions (Table 1) and assumptions using several examples that allow us to assess temporal and spatial variations of trait distributions. First, we searched the literature to determine if trait distributions measured from individuals actually do shift across local environmental gradients. Second, we assessed the dynamics of trait distributions and ecosystem carbon flux measures using data from an elevational gradient in Colorado. Third, we assessed the temporal dynamics of trait distributions and ecosystem net primary productivity using the Park Grass Experiment (PGE) from Rothamsted, UK. Lastly, to assess potential linkages with larger scale biogeographic gradients, we review recent studies that assess shifts in trait distributions across large-scale biogeographic environmental gradients.

We primarily focus on assemblage variation in one trait, SLA, because it appears to vary more than other traits in Eq. (11). Thus, we begin to assess predictions from TDT (Table 3) by substituting the mean SLA value for $\langle b_0 \rangle$. If other traits in Eq. (11) also vary or covary with each other across gradients, TDT would allow us to explore this as well. For example, a shift in the mean community carbon use efficiency (or c) will lead to a decrease in NPP as observed (DeLucia et al., 2007). Utilizing Eq. (11), we can now codify several additional TDT predictions based on SLA (see Table 3).

6.3 Shifts in Trait Distributions and Ecosystem Measures Across Local Abiotic Gradients

We tested several predictions generated by TDT (Table 1) with data collected along an elevational gradient in Colorado. We (Henderson, Sloat, and Enquist) have measured community composition, ecosystem fluxes, and traits of *all* individuals in several communities across an elevational gradient within subalpine communities near the Rocky Mountain Biological Laboratory (RMBL, Gunnison Co., CO, USA). Sites ranged from 2460 to 3380 m and had similar slope, aspect, and vegetation. The lowest elevation site is characterized as a semi-arid sagebrush scrub, whereas subalpine meadow communities dominate at the higher elevations. To estimate $C(z)$ leaf traits and biomass was measured from every individual within a 1.2×1.2 m plot. Measures of total ecosystem carbon production, community-weighted SLA, and variances were obtained by harvesting

biomass and measuring total ecosystem carbon fluxes or net ecosystem production (NEP; $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$). A more detailed listing of the methods used in our analyses is given in the [Appendix](#).

6.4 Local Tests of TDT Predictions

We next tested several predictions generated by TDT ([Table 1](#)) with data from the PGE from Rothamsted, UK. The PGE follows grassland plant community composition over a 140-year period. Started in 1956, it is the oldest ecological experiment in the world ([Silvertown et al., 2006](#)). The dataset is unique as it allows us to assess community responses to an environmental driver—the experimental altering of soil nutrient availability. We use this dataset to assess how a change in the environment, E , in this case soil nutrients, differentially influences community composition and ecosystem function via the trait distribution, $C(z)$. Within this experimental setup, the main environmental driver is a nutrient addition in the fertilized plot. We first focused on quantifying community SLA frequency distributions. However, as a more direct test of TDT, we also assessed two other key traits, plant height and seed size (see [Appendix](#)). To approximate the trait distribution, $C(z)$, we assigned species mean traits to species found within the PGE from the LEDA database ([Kleyer et al., 2008](#)). A detailed listing of the methods used in our analyses is given in the [Appendix](#) including background of the PGE.



7. RESULTS

7.1 Community Trait Shifts Across Local Gradients

Numerous studies have documented shifts in the traits of communities and assemblages across environmental gradients ([Ackerly, 2003](#); [Choler, 2005](#); [Fonseca et al., 2000](#); [Swenson and Enquist, 2007](#)). However, many studies generally calculate a species mean trait as part of a species list, thus ignoring intraspecific variation. In contrast, several recent studies have measured traits within communities to assess community-level trait shifts ([Albert et al., 2010](#); [Gaucherand and Lavorel, 2007](#); [Hulshof et al., 2013](#); [Lavorel et al., 2008](#)). For example, [Cornwell and Ackerly \(2009\)](#) show that across a gradient of water availability, the community mean and intraspecific mean SLA significantly shifted such that drier environments have lower mean SLA ([Fig. 4A](#)).

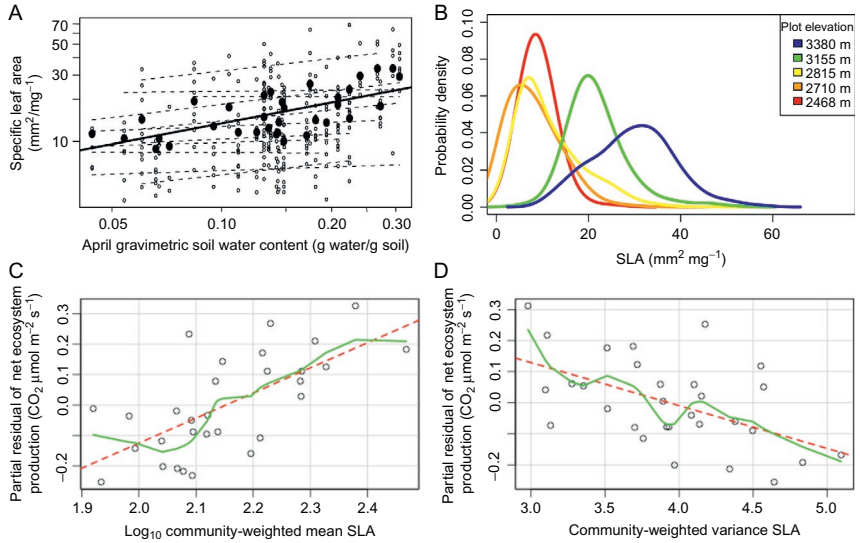


Figure 4 Two examples of shifts in community trait distributions across gradients. (A) Data from [Cornwell and Ackerly \(2009\)](#) showing shifts in the community mean and intraspecific mean value of the trait distribution for specific leaf area or SLA. Dashed lines represent least-squares fits for a given species and show the change in the population mean intraspecific variation across the gradient in soil water. Solid points and the solid line show the least-squares regression for the arithmetic mean community or interspecific value for SLA. *Note:* consistent with the assumption of a shift in an ‘optimal phenotype’ across environmental gradients, intraspecific trait shifts are in the same direction as the community or interspecific shift. (B) Data from our Colorado elevational gradient showing the probability density distributions of SLA based on all individuals in a 1.3×1.3 m plot at five sites along an elevational gradient. The number of individuals at each site is 2468 m = 234, 2710 m = 639, 2815 m = 938, 3155 m = 282, and 3380 m = 160. Across the elevational gradient, the community trait distribution of all individuals significantly shifts with changes in the mean community trait and variance. In (C) and (D), for the Colorado plots, we assess how changes in the community-weighted mean and variance of five plots within each site contributes to variation in net ecosystem production of CO₂ (NEP). These plots are partial residual plots showing linearization of NEP relationships with the community-weighted values of mean SLA and variance in SLA. As predicted by TDT, ecosystem carbon flux is positively related to a shift in mean SLA but negatively related to an increase in community trait variance. In contrast, variation in species diversity explains none of the variation NEP in this system.

7.2 Shifts of Community Trait Distributions Across Environmental Gradients

Data from our elevational gradient at the Rocky Mountain Biological Lab, Gothic, CO (see [Bryant et al., 2008](#); [Sides et al., 2014](#); [Sloat et al., 2014](#))

provide one of the first studies to measure the functional traits of every single individual within a given community (Fig. 4B). Few studies have fully documented the community trait distribution by measuring trait values from every individual within the community. In this system, increasing elevation is associated with a decrease in temperature and increase in precipitation, and these changes drive the observed increase in SLA with elevation (Sides et al., 2014). With increasing elevation, leaves have less structural durability, and lower life spans due to a shorter growing season but have higher photosynthetic rates (B.J. Enquist et al., unpublished data). According to TDT, the elevational trend in the SLA distribution is due to a shift in the optimum trait value based on temperature and water availability. This shift should also correspond with a shift in ecosystem functioning. Further, any change in the variance of the distribution will also have impacts on ecosystem functioning and the ability of the site to respond to future shifts in the environment.

Assessing shifts in inter- and intraspecific trait variation allows us to assess two central assumptions of TDT. First, trait distributions show directional shifts across gradients. According to Eq. (10), for a given E , those individuals with phenotypes that are closer to the mean community value should have, on average, the highest growth rates. Previous studies along this same gradient and study site have documented a rapid turnover of species with elevation (Bryant et al., 2008). Figure 4B indicates that the strong species diversity gradient is also reflected by a shift in traits. Note the range of trait variation is approximately 2–3 orders of magnitude. This range of trait values observed within communities in our gradient is approximately half of the fraction of the variation observed in SLA across the globe in all plants (Reich et al., 1997). So, across the span of ~ 25 km distance between their study sites, we observe a significant fraction of the trait variation that is observed within species across the globe. As more studies document shifts in SLA across strong environmental gradients (such as elevation, flooding, soil water availability, disturbance), it is becoming clear that the magnitude of change can be nearly as large the global variation in the trait (Elser et al., 2010; Violle et al., 2012). These results suggest that more local studies of community trait distributions are reasonable proxies or natural laboratories for scaling up trait-based ecology across large global climate gradients as well as to predict future climate change scenarios.

Second, analyses from Sides et al. (2014) and Cornwell and Ackerly (2009) also provide a key assessment to a core assumption of TDT. According to TDT, for a given environment, E , if there is a mean optimal phenotype that maximizes growth rate given an environmental trade-off

(Fig. 2), then external filters and/or selection/plasticity will then promote convergence of traits around this local optimal phenotype (Norberg et al., 2001; see also Violle et al., 2012). Both of these studies show that patterns of intraspecific mean trait shifts across an environmental gradient are in the *same* direction as the interspecific community shift across the gradient (see Fig. 4A). This is consistent with the expectation that either selection and/or phenotypic plasticity has resulted in individuals that adjust their phenotypes to better match an optimal phenotype within each community.

7.3 Shifts of Assemblage Trait Distributions Across Broad Environmental Gradients

Across broad-scale geographic gradients, recent geographic trait mapping analyses from Swenson et al. (2012) and Šímová et al. (2014) show that the mean assemblage trait value of many plant functional traits varies directionally across biogeographic scales. Geographic variation in the mean tree assemblage SLA as well as tree size (height, a proxy for plant mass, m) shows significant shifts in both traits across gradients at the biogeographic scale (Fig. 5). As is assumed in TDT, across a given environmental gradient, E , the mean community value, $C(z)$ or $C(b_0)$, will shift. Indeed, across North America, Šímová et al. find that the mean assemblage SLA is positively correlated with annual precipitation ($r^2 = 0.539$) but negatively related to annual temperature seasonality ($r^2 = -0.440$) (see Table 2 in Šímová et al., 2014).

7.4 Building Better Models for Variation in Ecosystem Function via the Shape of Trait Distributions and MST

At the local scale, measures of trait distributions associated with our theory in principle can be used to scale up to ecosystem function as well as to predict potential future community responses to climate change. Focusing on a key trait, SLA, and the total community biomass, C_{Tot} , we fit a simplified version of the TDT scaling model $\text{NEP} \propto \langle \text{CWM}_{\text{SLA}} \rangle \cdot \langle \text{CWV}_{\text{SLA}} \rangle \cdot C_{\text{Tot}}^b$, where NEP is the Net Ecosystem Production ($\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$). To focus on just the effects of SLA and C_{Tot} on NEP, we allowed each sample site along the gradient to be a random factor in the model. Here, b is the fitted exponent. Here, the values $\langle \text{CWM}_{\text{SLA}} \rangle$ and $\langle \text{CWV}_{\text{SLA}} \rangle$ are the community abundance-weighted mean and variance in SLA, respectively, based on log transformed trait values. The fitted model explains $\sim 71\%$ of the variation in NEP ($\text{df} = 22$, $F = 11.04$, $p < 0.0001$, $\text{AIC} = -24.39$). In support of TDT,

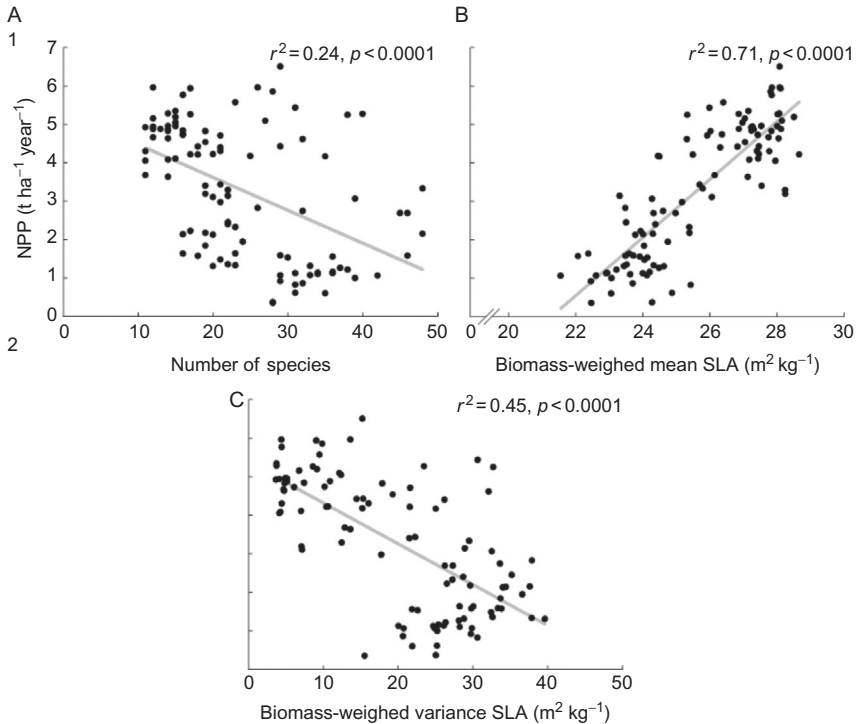


Figure 5 Correlations between annual net primary productivity (NPP) and (A) species diversity per plot; (B) the plot community biomass-weighted mean, CWM, of the SLA distribution; and (C) the plot community biomass-weighted variance, CWV, of the SLA distribution for the four selected plots from the Park Grass experiment. Each dot represents the annual aboveground biomass production of a given plot in a given year. Compare these relationships to the predictions with Trait Driver Theory and Biodiversity and Ecosystem Functioning Theory (Table 3). Note that there is a negative correlation between NPP and diversity, while the opposite is predicted by classical Biodiversity–Ecosystem Functioning theory. However, in accordance with TDT, there is a *negative* relationship between the variance of the abundance-weighted community trait distribution and a *positive* relationship between mean-weighted trait and NPP. These results show a direct linkage between a critical trait that influences plant growth, forces that influence the shape of the trait distribution, and how both changes in trait mean and variance then shape ecosystem functioning.

both increases in $\langle \text{CWV}_{\text{SLA}} \rangle$ and C_{Tot} each increase community NEP ($p=0.023$, $t=0.337$; and $p=0.058$, $t=2.332$ respectively) while increases in $\langle \text{CWV}_{\text{SLA}} \rangle$ tends to *decrease* NEP ($p=0.068$) (see Fig. 4). The fitted exponent for b is different than the predicted value of $3/5$ ($b=0.13 \pm 0.14$) but the range of variation in plot biomass, C_{Tot} in this study is generally less than an order of magnitude. In contrast, variation in species diversity explains

none of the variation in NEP in this system either as a single predictor ($r^2 = 0.029$, $p = 0.3617$). Allowing each sample site along the gradient to be a random factor in the model shows that species richness has a negative effect on NEP in the model ($t = 0.01$, $p = 0.03$). Additional analyses underscore the importance of both $\langle \text{CWM}_{\text{SLA}} \rangle$ and $\langle \text{CWV}_{\text{SLA}} \rangle$ on influencing variation NEP across this elevational gradient (see [Appendix](#)). Together, these results support several key predictions of TDT—the shift in the mean of $C(b_0)$ is closely tied to environmental drivers and that shifts in the mean and variance of $C(b_0)$ are a primary driver of variation in community carbon flux ([Fig. 4C and D](#)).

Across broad climatic gradients, recently [Michaletz et al. \(2014\)](#) utilized MST to predict variation in annual net primary productivity (NPP, grams of biomass per area per year). In support of MST, rates of growing season NPP scaled with total autotrophic biomass indistinguishable from the allometrically ideal value of 3/5 predicted value in [Eq. \(12\)](#). Their analysis support another prediction of TDT that controlling for scaled effects of total biomass, C_{Tot} , and stand age on NPP shows that shifts in $\langle b_0 \rangle$, primarily because of shifts in $\langle \text{SLA} \rangle$, will also shift variation in NPP ([Fig. 7](#)).

At larger biogeographic scales, assemblage trait maps such as [Fig. 5](#) could then be used to predict ecosystem functioning. At these larger geographic scales, if the distribution of SLA still reasonably approximates rates of biomass production, then, according to TDT, regions with high mean SLA and low variance should have the highest rates of instantaneous net primary production. In general, recent compilation of geographic variation in instantaneous rates of terrestrial ecosystem NPP from remotely sensed data indicates that areas with the highest instantaneous rates of NPP do generally correspond² to assemblages with high mean and low variance in SLA. However, according to [Eq. \(16\)](#), one should also control for total system biomass (which correlates with variation in tree height) as well as variation in the other traits that also can influence NPP. Nonetheless, the correspondence between biogeographic variation traits and predictions from TDT is a promising future direction.

7.5 Temporal Trait Shifts Across Fertilization Gradients

The results from the long-term dynamics and fertilization experiment from Rothamsted are given in [Tables 2 and 3](#) as well as in [Figs. 6 and A3–A5](#). Within the Rothamsted dataset, all of the traits studied, the biomass–

² http://daac.ornl.gov/NPP/npp_home.shtml#.

Table 2 Observed Temporal Changes in the Central Moments of the Community Trait Distribution $C(z)$ in the Park Grass Experiment (see Figures A3–5 in Appendix)

	Moment of Community Trait Biomass Distribution, $C(z)$	Control		Corresponding TDT Predictions in Table 1
		(Unfertilized)	Fertilized	
SLA	Mean (CWM)	0.65*** (–)	0.40** (+)	See I(a) and I(b)
	Variance (CWV)	0.48** (+)	0.26* (–)	See II(a)–(e)
	Skewness (CWS)	0.13 ^{ns.}	0.29* (–)	See III(a)–(b)
	Kurtosis (CWK)	0.69*** (–)	0.10 ^{ns.}	See IV(a)–(b)
Height	Mean (CWM)	0.61** (–)	0.04 ^{ns.}	See I(a) and I(b)
	Variance (CWV)	0.27* (–)	0.51** (+)	See II(a)–(e)
	Skewness (CWS)	0.17 ^{ns.}	0.02 ^{ns.}	See III(a)–(b)
	Kurtosis (CWK)	0.38** (+)	0.20 ^{ns.}	See IV(a)–(b)
Seed mass	Mean (CWM)	0.21 ^{ns.}	0.01 ^{ns.}	See I(a) and I(b)
	Variance (CWV)	0.38** (–)	0.01 ^{ns.}	See II(a)–(e)
	Skewness (CWS)	0.56** (–)	0.00 ^{ns.}	See III(a)–(b)
	Kurtosis (CWK)	0.31* (–)	0.01 ^{ns.}	See IV(a)–(b)

Predictions from Trait Driver Theory correspond to the cells indicated in Table 1. As an estimate of the central moments of $C(z)$, we estimated the community-weighted values for the mean, variance, skewness, and kurtosis (CWM, CWV, CWS, and CWK, respectively). Values are the Pearson product–moment correlations, r , between time since the start of the experiment and the corresponding trait distribution moment for the three main traits investigated, specific leaf area or SLA, adult height, and seed size. In accordance with TDT, for the trait SLA, the distributions of the fertilized and unfertilized plots have diverged for all trait moments. Further, in accordance with TDT, increasing fertilization leads to an increasing skew of the SLA distribution, but not for seed size and reproductive height, traits that do not directly underlie the growth equation, indicating that fertilization is a strong environmental driver that influences plant growth. Correlations are for both unfertilized: plot 2 and fertilized: plot 16. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$; ns., not significant; (–), negative relationship; (+), positive relationship. Observed shifts in the central moments of SLA are generally in accordance to predictions from TDT (see Table 1 and text for details).

weighted distribution of the trait SLA was associated with the most prominent shifts in the central moments of communities in response to fertilization (Fig. A1; Table 2; see also Appendix). For both fertilized and control plots, all four moments of the community SLA distribution changed significantly. The overall effect of fertilization on the community trait

Table 3 Correlations Between Trait Moments and Annual Net Primary Productivity for the Park Grass experiment

	Central Moment of Community Trait Distribution	Observed Correlations	Predicted Response–Trait Driver Theory	Predicted Response–Biodiversity Theory
SLA	Mean (CWM)	0.71*** (+)	+	NP
	Variance (CWV)	0.45*** (–)	–	+
	Skewness (CWS)	0.28*** (+)	NP	NP
	Kurtosis (CWK)	0.19*** (+)	+	–

Also listed are the predicted signs of the correlations made from Trait Driver Theory (see text and Table 1) and from Biodiversity–Ecosystem Functioning theory (assuming the theory of Tilman et al., 1997 where variability in trait SLA is a good proxy for variation in species richness via niche or trait space). *Note:* both theories make opposite predictions for the signs of the correlations. NP, no specific prediction is made as such predictions would depend upon specifics of system. As an estimate of the central moments of $C(z)$, we estimated the community biomass-weighted values for the mean, variance, skewness, and kurtosis (CWM, CWV, CWS, and CWK, respectively).

*** $p < 0.001$.

(–), Negative relationship; (+), positive relationship.

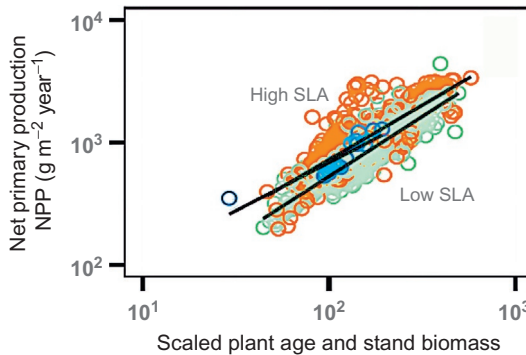


Figure 6 In support of Trait Driver Theory, woody plant net primary production scales with total assemblage biomass (see Eqn. 12). Further, for a given amount of biomass, NPP will be modified by shifts in the mean community leaf trait, specific leaf area or SLA (see Eqn. 15). *Figure modified from Michaletz et al. (2014).*

distribution is consistent with fertilization differentially favouring certain phenotypes (Chapin and Shaver, 1985; Suding et al., 2005; Tilman, 1982) and a replacement of slower growing species with faster growing species (Chapin, 1980; Grime and Hunt, 1975; Knops and Reinhart, 2000). Similar to past findings, across all plots, the community mean SLA increased (see Craine et al., 2001; Knops and Reinhart, 2000) and the variance

decreased. The directional community trait shift is reflected in increased skewness values (differing from zero). Fertilization also led to a shift in the kurtosis but only for SLA values. Specifically, SLA shifted from negative kurtosis values to zero or positive kurtosis values, suggesting that fertilization increased rates of competitive exclusion of suboptimal trait values leading to a more peaked trait distribution. Intriguingly, the direction and rate of change for fertilized versus control plots differed in sign and magnitude indicating that the trait distributions of control and experimental plots steadily diverged over time.

The PGE supports several predictions from TDT. First, TDT predicts that a shift in an environmental driver (fertilization in this case) should primarily be seen as a shift in traits associated with growth rate. Of all of the traits assessed, SLA showed the strongest shifts over time (Table 1). The other traits showed relatively little to no change over time. The disproportionate shift in SLA is consistent to expectations from TDT as SLA is the only trait directly linked to the growth function, $f(b_0)$. Second, consistent with a shifting in an optimal phenotype, the skewness of the fertilized plot increased over time. Third, consistent with a productivity–trait variance trade-off, the annual net primary productivity (NPP) was positively correlated with community mean and kurtosis of the SLA distribution but negatively correlated with variance in SLA (Fig. 6; Table 3). Lastly, shifts in the trait distribution are more closely tied to NPP than species richness (see Table 3). We observe a weak to negative correlation between species richness and NPP (Fig. 6A; Table A1). Together, our approach provides: (i) a more mechanistic understanding of the long-term response of the species and communities found within the PGE experiment and (ii) an alternative trait-based approach that can in principle be integrated with past ‘species richness based’ theories invoked to explain the decrease species richness with fertilization (Tilman, 1982).



8. DISCUSSION

We have shown that TDT can formalize numerous assumptions and approaches in trait-based ecology. We provide examples of how this can be done for several different biodiversity hypotheses in terms of the dispersion of traits (Tables 1 and 3). We further argue that ecological theories need to move beyond species richness and be recast in terms of organismal performance via functional traits. As a result, TDT offers an alternative framework to the standard taxonomic approach for linking biodiversity and ecosystem

functioning, where primacy has been placed on the importance of species richness. TDT instead focuses on the importance of ‘trait diversity’ via the shape of the trait distribution of individuals and shared performance currencies (e.g. growth). Because TDT incorporates intraspecific variation, it necessarily includes natural selection as a process that shapes the trait distribution. By incorporating interspecific trait variation, it also includes ‘selective’ processes at higher levels of organization within the community, such as species sorting (see also Shipley, 2010). We show that using TDT to analyze these processes leads to several predictions opposite to predictions made by Biodiversity–Ecosystem Function theory (Naeem and Wright, 2003; Tilman et al., 1997; see also Table 3); it also offers a useful alternative hypothesis by which to assess the linkage between ‘diversity’ (whether measured by species richness or via the trait distribution) and ecosystem functioning.

TDT also offers a predictive framework for management. Increasingly, trait-based approaches to management have shown that a focus on trait shifts due to land use, as well as management and agricultural practice, can yield deeper insight into the processes of concern to managers (Garnier and Navas, 2012). For example, biomass production, the timings of peak production and plant digestibility, and response time to disturbance can be predicted from the shape of the community trait distribution as well as many of the plant traits underlying our general growth equation (see studies and references listed in Garnier and Navas, 2012). It is intriguing to note that TDT predicts a trade-off between short-term productivity and long-term response to the environment. Agriculture focused on maximizing short-term productivity reduces trait variance by planting genetically homogeneous monocultures.

Our analyses find that none of the central moments of the trait distribution in the Park Grass dataset are correlated with species richness (see Table A1 in the Appendix). Indeed, species richness does not appear to be a reliable proxy for how the diversity of phenotypes and trait distributions respond to environmental change. Further, across large biogeographic gradients, recent studies have found the trait variance and the total functional trait space is often unrelated to species richness (Lamanna et al., 2014; Safi et al., 2011; Šímová et al., 2014; Fig. 7). The potential for improved predictions using trait and size distributions that can be linked to metabolic scaling fundamentally comes down to the increase in information contained in traits that is not necessarily present in a species-richness-based approach (Tilman et al., 1997) or even a phylogenetic approach to community ecology

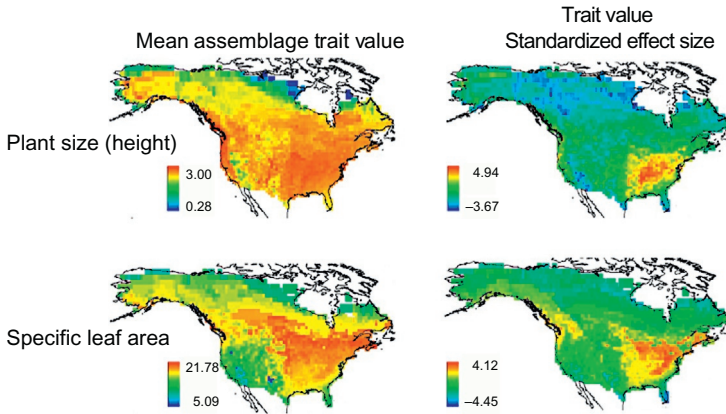


Figure 7 Biogeographic variation in the (A) mean species assemblage functional trait specific leaf area (SLA, $\text{m}^2 \text{g}^{-1}$) and plant height (m) and (B) the standardized effect size (SES) of the mean trait values across woody plant communities across North America. The SES measures if the observed value is significantly higher or lower than expected given the species richness. Thus, high SES values indicate greater mean trait values than expected from the observed number of species and low SES values indicate lower mean values than expected. The mean SLA is lowest in the low elevation-latitude tropical forests and tends to be highest in temperate forest and grassland regions. Across broad geographic gradients, both the mean and the variance significantly vary. According to Trait Driver Theory, areas that correspond to high mean SLA and larger standing stocks of biomass (corresponding to taller mean heights) should have the highest rates of instantaneous net primary production, NPP. However, NPP will also be modified by the assemblage variance in SLA. *Figure modified from Šímová et al. (2014).*

(Cavender-Bares et al., 2009). Although TDT is based on simplifying assumptions, it helps to better connect and scale trait-based ecology and MST with large-scale ecology and biogeography. It integrates and builds upon prior work that developed highly mechanistic trait-based models (Norberg et al., 2001; Zhang et al., 2013) and other work that predicted how individual growth rates change across scale with size and temperature (Enquist et al., 2007a). TDT thus enables ecological theories to be ‘scaled up’ to predict and test the consequences of how organismal response to climate change will ramify at the community and ecosystem levels across both local- and large-scale gradients in geography (space) and fluctuations in time (climate change).

It is becoming clear that multiple assembly processes—abiotic filtering, biological enemies, competition, facilitation, below-ground competition—likely operate simultaneously and at differing scales to structure communities and larger scale species assemblages (Cavender-Bares et al., 2009; Grime,

2006; Mayfield and Levine, 2010; Swenson and Enquist, 2009). The result is that the distribution of some traits may be more over- or under-dispersed than others. Future work should better link empirical data with theory to test the community and ecosystem responses when multiple trait drivers influence trait variance and when trait optima are strongly influenced by differing levels of ecological interactions (competition, predation, mutualism, etc.).

Our work has primarily focused on the traits and environmental drivers that underlie growth rate. Indeed, refining and extending TDT will also require better identification of the above- and below-ground traits that influence growth rate. Lastly, analysis of the shift in the mean and variance of assemblage values of many plant functional traits as well as stand biomass across biogeographic scales may provide the necessary basis to predict ecosystem functioning across large scales. Focusing on variation in SLA, TDT predicts that the distribution of SLA will influence rates of biomass production. Correcting for the effects of stand biomass (see Eq. 8), assemblages with high mean SLA and low variance should have the highest rates of instantaneous net primary production. In general, recent compilation of geographic variation in instantaneous rates of NPP from remotely sensed data indicates that areas with the highest instantaneous rates of NPP generally do correspond (Zhao and Running, 2010; Zhao et al., 2005) to assemblages with high mean and low variance in SLA regions identified by Swenson et al. (2012) and Šimová et al. (2014). TDT also predicts that while regions with relatively lower trait variance will be more sensitive to rapid directional climate change, assemblages with greater variance, however, would be expected to more closely track climate change. Future tests of TDT at the scale of global ecology should more formally assess the predictions of TDT at this scale by assessing the specific relationships between the trait distribution, vegetation biomass, and possible covariation of other traits.

The TDT prediction of an inverse relationship between trait variance and production is not necessarily in conflict with either ‘positive species complementarity’—niche partitioning allows species to capture more resources in ways that are complementary in both space and time (Tilman, 1999)—or ‘transgressive overyielding’ where species use resources in ways that are complementary in space or time to stably coexist with one another so that more diverse communities capture a greater fraction of available resources and produce more biomass than even their most productive species (see Tilman et al., 1997). TDT needs to be reconciled with these ideas because recent studies confirmed that within biodiversity experiments,

‘positive species complementarity’ does enhance ecosystem productivity (Cardinale et al., 2007). Given TDT, a natural question is how are trait distributions modified when complementarity effects are strong? Effects such as complementarity can be incorporated into TDT growth functions as explained in Savage et al. (2007).

In sum, we have argued that more powerful tests of biodiversity theories need to move beyond species richness and explicitly focus on mechanisms generating diversity via trait composition and diversity via the shape of trait distributions. The rise of trait-based ecology has led to an increased focus on the distribution and dynamics of traits across broad geographic and climatic gradients and how these distributions influence ecosystem function. However, a trait-based ecology that is explicitly formulated to apply across different scales (e.g. species that differ in size) and gradients (e.g. environmental temperatures) has yet to be articulated. The TDT presented here is a formalization of essential steps for mechanistically linking and scaling functional traits for individuals with the dynamics of ecological communities and ecosystem functioning. This TDT approach builds upon and complements existing trait-based approaches in ecology (e.g. Grime, 1998; Kraft et al., 2008; Lavorel and Garnier, 2002; Suding et al., 2008a). It is appealing because it can connect individual physiology and traits with ecosystem dynamics and how both respond to climate change (Suding et al., 2008b), geographic gradients, and differing ecological processes (e.g. niche vs. neutral; see Weiher et al., 2011). Given the increasing ability to remotely sense numerous traits of terrestrial vegetation (Asner et al., 2014; Doughty et al., 2011) and the increasing access to both plant and animal trait data (Kattge et al., 2011), TDT and its elaborations are ripe for providing empirically grounded, mechanistic models of ecosystem dynamics from local to large scales.

ACKNOWLEDGEMENTS

We thank Mark Westoby who provided enthusiasm and input during the writing of initial drafts of this chapter. We also thank the ARC-NZ Vegetation Network, working group 36, led by V.M.S., for their generosity and support in bringing us all together to meet and to initiate this collaboration and chapter. We thank other members of working group 36, especially Christine Lamanna, Tony Dell, Mick McCarthy, and Graham D. Farquhar, who helped us solidify our central arguments. C.V. was supported by a Marie Curie International Outgoing Fellowship within the 7th European Community Framework Program (DiversiTraits project, no. 221060). B.J.E. and V.M.S. were supported by an NSF ATB award and B.J.E. by an NSF Macrosystems award. C.T.W. was supported by NSF Grant DEB-0618097. J.N. was supported by the Swedish Research Council and FORMAS Grant EKOKLIM. Lastly, we thank the Park Grass staff, in particular Andy Macdonald and Margaret Glendining, for assistance with and making available the Park Grass data.



APPENDIX

A.1 Using TDT to Recast Ecological Hypotheses

A unique attribute of TDT is that several differing ecological theories can now be recast in terms of how they influence the shape of trait frequency distributions ($C(z)$). Each of these theories makes differing hypotheses that influence the shape of trait distributions and the functioning of ecosystems:

- (i) *H1: Phenotype–environment matching*: This hypothesis states that species are more successful in different parts of the landscape because individuals have different trait values across space that are better adapted to local features of that space, such that the mean ‘phenotype’ matches variation in the local environment (Westoby and Wright, 2006). This distinction builds upon observations that stem back to Schimper (1898, 1903) that form the foundation for understanding changes in fitness and functional traits and species composition across gradients (Levins, 1968; Westoby and Wright, 2006). A prediction of this hypothesis is that, either due to convergent evolution or abiotic filtering of relevant traits, similar environments should be dominated by species with similar trait values (see discussion in Karr and James, 1975; Mooney, 1977; Orians and Paine, 1983).
- (ii) *H2: The competitive-ability hierarchy hypothesis*: This hypothesis states that the strength of competition between individuals is driven by the distance between individuals as measured according to their functional traits (Freckleton and Watkinson, 2001). The competitive-ability hierarchy hypothesis leads to opposite predictions than the niche-based competition–trait similarity and competition–relatedness hypotheses (see below; Mayfield and Levine, 2010). Here, the resulting competitive hierarchy will cause a reduction in the trait variance over time and increased functional clustering because individuals that share a trait value will outcompete individuals with different trait values (Freckleton and Watkinson, 2001; Kunstler et al., 2012; Mayfield and Levine, 2010).
- (iii) *H3: Abiotic filtering*: The importance of local abiotic forces is reflected in the community trait range and variance. Abiotic filtering hypothesis states that increasingly more stressful environments will limit the range and variance influence. This hypothesis was formalized by Keddy and colleagues (Keddy, 1992; Kraft and Ackerly, 2010; Weiher and Keddy, 1999). Similarly, on ecological time scales, due to phenotype–

environment matching, increasingly more stressful environments, E , will increasingly restrict the range of trait values that could co-occur within a given environment. This ‘trait filtering hypothesis’ states that the abiotic environment filters trait values so as to limit the variance and range of the trait distribution. This hypothesis, which can be seen as an ecological scale version of the ‘favourability’ hypothesis of Terborgh (1973), predicts that more physiologically stressful environments (frost, high salinity, drought, etc.) should place especially rigid filters on the types of phenotypes (i.e. traits) that can survive and potentially co-occur (e.g. Kraft et al., 2008). Note, as discussed in (ii) and (iii), the variance and range of a trait distribution $C(z)$ are also influenced by biotic forces. Similarly, repeated disturbance or environmental variability may minimize local interactions and could also increase community trait variance (Grime, 2006).

- (iv) *H4: Strength of local biotic forces is revealed via trait variance and kurtosis:* Differing biotic community assembly hypotheses can differentially influence the spacing of trait values within the range of filtered phenotypes. As stated in (ii), competition for a common limiting resource would ultimately lead to competitive exclusion (e.g. see Tilman, 1982) resulting in a convergence of ‘superior competitor’ phenotypes (Abrams and Chen, 2002; Mayfield and Levine, 2010; Savage et al., 2007). This convergence would be reflected by decreasing variance and an increase in ‘peakedness’ of the trait distribution or an increase in positive kurtosis (Navas and Violle, 2009). In contrast, according to Chesson (2000), if traits map onto niche differences, increased niche (trait) differentiation will lead to increasing coexistence of individuals with differing traits. These classical niche partitioning models predict that competition will limit functional (trait) similarity (MacArthur and Levins, 1967) and thus increase in the spacing between co-occurring phenotypes (see Diamond, 1975; MacArthur, 1958). Similarly, biological enemies (Kraft and Ackerly, 2010), facilitation (Brooker et al., 2008), and frequent disturbance (Grime, 1998) can maintain trait diversity (e.g. an over-dispersion of phenotypes). Niche packing models result in either a broader or evenly dispersed trait distribution (high variance) or even a multimodal trait (negative kurtosis) distribution.
- (v) *H5: Assessing neutral forces via the shape of local and regional trait distributions:* An alternative hypothesis to (ii) and (iii) is that local communities are primarily structured by stochastic dispersal, drift, and dispersal limitation (Hubbell, 2001). Such a neutral scenario would predict on average, for traits not associated with dispersal, little to no difference

in the shape of the community trait distribution when sampled across differing spatial scales. Further, for traits not associated with dispersal ability, there should be no relationship between trait distribution and changes in the environment.

A.2 Integrating Metabolic Scaling Theory into TDT

Within TDT, the total biomass associated with a trait, z , is denoted by $C(z)$. This notation avoids ever needing to account for individual mass, M , or number of individuals with mass. In contrast, the growth equations Metabolic Scaling Theory are phrased in terms of individual mass, M , where organismal growth rate, dM/dt , is given by

$$\frac{dM}{dt} = b_0(z)M^{3/4} \quad (\text{A1})$$

where $b_0(z)$ is a coefficient that depends on a single or set (meaning z is a vector) of traits.

To integrate MST and TDT, we first recognize that, $C(z)$, the biomass associated with trait z can be expressed as $C(z) = \int dM C(z, M) = \int dM N(z, M)M$, where $C(z, M)$ is the mass density of individuals with both trait value z and individual mass M , while $N(z, M)$ is the number density of individuals that have both trait value z and individual mass M . In this expression, we have integrated over all possible values of mass, M , so we have the total biomass of *all* individuals with trait z . Furthermore, note that integrating this over all traits, z , gives the total biomass, $C_{\text{Tot}} = \int dz C(z) = \int dz \int dM N(z, M)M$. Consequently, we can multiply both sides of Eq. (A1) by $N(z, M)$, integrate both sides over $\int dz \int dM$, and multiply and divide the right side by C_{Tot} to obtain

$$\int dz \int dM N(z, M) \frac{dM}{dt} = \left[\frac{\int dz \int dM N(z, M) b_0(z) M^{3/4}}{\int dz \int dM N(z, M) M} \right] C_{\text{Tot}} \quad (\text{A2})$$

At steady state, $N(z, M)$ is not changing in time, so we can move it inside of the derivative with respect to time, and we can also move the integrals inside of the derivative because the integration over all possible traits and masses is not a time-dependent object

$$\frac{d \left[\int dz \int dM N(z, M) M \right]}{dt} = \frac{dC_{\text{Tot}}}{dt} \quad (\text{A3})$$

Furthermore, we can express the bracketed term on the right side of Eq. (A1) as a mass average of a growth function as follows

$$\left[\frac{\int dz \int dMN(z, M) M \left[b_0(z) M^{-1/4} \right]}{\int dz \int dMN(z, M) M} \right] = \left[\frac{\int dz \int dMC(z, M) \left[b_0(z) M^{-1/4} \right]}{\int dz \int dMC(z, M) M} \right] = \left\langle b_0(z) M^{-1/4} \right\rangle_C \quad (\text{A4})$$

where the C subscript denotes that the average is taken with respect to the biomass. Combining all of this, we obtain the equation for the scaling of Net Primary Productivity or NPP equation

$$\frac{dC_{\text{Tot}}}{dt} = \left\langle b_0(z) M^{-1/4} \right\rangle_C C_{\text{Tot}} \quad (\text{A5})$$

This equation is in the most generic form of a TDT equation, and the growth function $f(z) = b_0(z) M^{-1/4}$ can be expanded such that the biomass growth equation can be expressed in terms of the biomass-weighted central moments of the trait z , such as the variance, skewness, and kurtosis.

We now consider a few special cases of Eq. (A5) to relate to the TDT and scaling equations already in the literature. In the case that there is only a single mass value, M^* , or a very small range of mass values, the number density becomes $N(z, M) = N(z) \delta(M - M^*)$ in terms of a Dirac-delta function for the mass dependence. Therefore, $\int dz \int dMN(z) \delta(M - M^*) M \left[b_0(z) M^{-1/4} \right] = \int dz N(z) M^* \left[b_0(z) (M^*)^{-1/4} \right] = (M^*)^{-1/4} \int dz C(z) b_0(z)$, and Eq. (A4) becomes

$$(M^*)^{-1/4} \frac{\int dz C(z) b_0(z)}{\int dz C(z)} = (M^*)^{-1/4} \left\langle b_0(z) \right\rangle_C \quad (\text{A6})$$

so the scaling of NPP equation becomes

$$\frac{dC_{\text{Tot}}}{dt} = (M^*)^{-1/4} \left\langle b_0(z) \right\rangle_C C_{\text{Tot}} \quad (\text{A7})$$

The term $(M^*)^{-1/4}$ can be thought of as an overall normalization to the growth function $f(z)$ from TDT. As such, this result reveals that TDT, as

originally formulated, essentially ignores individual mass. Thus, based on Eq. (A1), growth functions within TDT should have a roughly $(M^*)^{-1/4}$ hidden with the normalization constant for their growth function.

Conversely, as a special case, we consider the function $b_0(z)$ to be a constant b_0 that occurs when $z = z^*$. In this case, $N(z, M) = N(M)\delta(z - z^*)$ and $\int dz \int dMN(M)\delta(z - z^*)M[b_0(z)M^{-1/4}] = b_0 \int dMN(M)M^{1/4}$, and Eq. (A4) becomes $b_0 \int dMN(M)M^{3/4}/C_{\text{Tot}}$. Combining these terms gives the scaling of NPP equation

$$\frac{dC_{\text{Tot}}}{dt} = \frac{b_0 \int dMN(M)M^{3/4}}{C_{\text{Tot}}} C_{\text{Tot}} = b_0 \int dMN(M)M^{3/4} \quad (\text{A8})$$

Following the arguments in [Enquist et al. \(2009\)](#), we can substitute $N(M) \propto M^{-11/8}$ to obtain $M_b \propto C_{\text{Tot}}^{8/5}$ where the subscript denotes that largest mass in the group. Using these relationships

$$\frac{dC_{\text{Tot}}}{dt} \propto b_0 \int dMM^{-11/8}M^{3/4} \propto b_0 \int dMM^{-5/8} \propto b_0 M_b^{3/8} \propto b_0 C_{\text{Tot}}^{3/5} \quad (\text{A9})$$

Defining a new constant b'_0 to denote the product of b_0 with all of the proportionality constants, the overall scaling of NPP equation becomes

$$\frac{dC_{\text{Tot}}}{dt} = b'_0 C_{\text{Tot}}^{3/5} \quad (\text{A10})$$

in accord with the Net Primary Productivity scaling equation derived by [Enquist et al. \(2009\)](#).

As a final special case, we consider the trait z and the mass M to be uncorrelated and the number density to be a separable function such that $N(z, M) = N(M)N(z)$. Therefore, using results from Eq. (A9) and the definition of an average, Eq. (A4) can be expressed as

$$\frac{\int dz N(z) b_0(z) \int dMN(M)M^{3/4}}{\int dz N(z) \int dMN(M)M} = k \langle b_0(z) \rangle C_{\text{Tot}}^{-2/5} \quad (\text{A11})$$

where k captures the proportionality constants in deriving Eq. (A5). Substituting this into our overall growth equation (Eq. A5) yields

$$\frac{dC_{\text{Tot}}}{dt} = k \langle b_0(z) \rangle C_{\text{Tot}}^{3/5} \quad (\text{A12})$$

where the average is now the standard abundance average and is *not* the biomass-weighted average. This is the growth equation in the scaling form for this special case. Alternatively, for this special case, we could express Eq. (A4) as

$$\frac{\int dz N(z) b_0(z) \int dMN(M) M [M^{-1/4}]}{\int dz N(z) \int dMN(M) M} = k \langle b_0(z) \rangle \langle M^{-1/4} \rangle_C \quad (\text{A13})$$

and the NPP scaling equation becomes

$$\frac{dC_{\text{Tot}}}{dt} = k \langle b_0(z) \rangle \langle M^{-1/4} \rangle_C C_{\text{Tot}} \quad (\text{A14})$$

This equation is completely equivalent to Eq. (A13) but expresses the growth function more in terms of the TDT framework such that the right side appears to have an overall linear dependence in C_{Tot} , and as a result, we have a mixture of types of averages, with the function $b_0(z)$ being abundance averaged and the $M^{-1/4}$ being biomass averaged.

The major results of this section are Eq. (A5), which is the most general formulation of the growth equation because it does not rely on traits or mass being constant or uncorrelated. In this form, the growth equation is like the TDT formulation, but as the special cases below it reveal, $\langle b_0(z) M^{-1/4} \rangle_C$ may hide extra dependencies on C_{Tot} . Equation (A7) is the result when the mass is constant and is expressed in the form of TDT equations such that it is linear in C_{Tot} and reveals an overall $M^{-1/4}$ for adjusting the growth function across groups. Equation (A10) is the special case where the traits are constant and reduces to the exact scaling equation given in Enquist et al. (2009). Finally, when traits and mass are uncorrelated, Eqs. (A13) and (A14) are two different but completely equivalent ways to express the growth function. Equation (A13) is in the form of scaling equations by consolidating the mass average with the C_{Tot} dependence, while Eq. (A14) is in the form of TDT equations by keeping two averages around, including one that is an abundance average and one that is a biomass-weighted average. For all of these equations and cases, the functions inside the averages can be expanded in terms of moments as done for TDT for biomass-weighted averages or as done in 2004 for abundance-weighted averages.

A.3 Growth Functions Across Environmental Gradients: Incorporating Trade-Offs into TDT

Importantly, as discussed in the main text, Eq. (9) predicts an unbounded growth response such that increasingly larger values of b_0 always leads to increased growth, which realistically cannot continue indefinitely. A key assumption of TDT is that there is fundamental trade-off between a given trait value and the performance of an organism across an environmental gradient, E . The final step to integrate a general TDT that can link traits, organismal performance, and environmental gradients is to specify trade-offs between underlying traits, growth, and metabolic scaling.

Within a given environment, E , an important question is what would prevent the average metabolic normalization, $\langle b_0 \rangle$, from becoming infinitely big or small? In the case of growth rate, possible trade-offs likely include the types of limiting resources individuals use or the environmental conditions for optimal growth. So, individuals that allocate internal resources to specific traits defined by b_0 may reduce the impact of one limiting environmental factor, but this would necessarily incur a disadvantage with respect to another environmental limiting factor.

A trade-off or cost function can be formulated within the growth function, f . Multiplying this cost function by f shows that, for a given E , the growth function has a maximum at z_{opt} or here $b_{0,\text{opt}}$, and as a result, the second derivative of f (the second term in Eqs. A5 and A14) will be negative, as long as $\langle b_0 \rangle$ is close to $b_{0,\text{opt}}$.

We can specify a generic form of a trade-off by following [Norberg et al. \(2001\)](#). We can approximate a trade-off by first invoking a general quadratic or Gaussian cost function on the community value ($\langle b_0 \rangle - b_{0,\text{opt}}$). We add a cost function to Eqs. (A5) and (A14). This provides a general form of a trade-off. That new cost function that is multiplied by f could be a general quadratic $\left[1 - \left(\frac{\langle b_0 \rangle - b_{0,\text{opt}}}{\sigma^2} \right)^2 \right]$ or Gaussian function, $\exp \left[- \left(\frac{\langle b_0 \rangle - b_{0,\text{opt}}}{\sigma^2} \right)^2 \right]$. Here, σ^2 is the observed standard deviation in the trait or b_0 observed within the assemblage. Both cost functions reduce to 1 when $\langle b_0 \rangle - b_{0,\text{opt}}$ (e.g. for a given environment E , the observed mean community trait, z_{opt} , and average metabolic normalization, $\langle b_0 \rangle$, are at the local optimum). Note both decrease in value as you go away from $b_0 - b_{0,\text{opt}}$. Dividing through by σ^2 defines the penalty for individual growth rate for being away from the optimum. Thus, for a given environment, E , characterized by a unique $b_{0,\text{opt}}$, the growth function can be made more explicit in terms of a generic trade-off where

$$f(b_0) = c_0 b_0 \langle M^{\theta-1}(b_0) \rangle \left(1 - \frac{(b_0 - b_{0,\text{opt}})^2}{\sigma^2 b_0} \right) \quad (\text{A15})$$

This is a modified growth function and is characterized in Fig. 2. It can be made more specific by incorporating the traits that then define b_0 . Here, the first term is the general growth equation from the relative growth rate literature (Poorter, 1989) that has been more formally derived in metabolic scaling theory. The second term of Eq. (A1) is the associated trade-off. As a result, the second term in the TDT Eq. (5) in the main text gives how much of whole-community biomass production is *reduced* due to the amount of trait variance, V , in the community because of the explicit trade-off function, the second derivative of f , $\frac{d^2 f_{b_0 = \langle b_0 \rangle}}{db_0^2}$, would then be negative near $b_{0,\text{opt}}$. As a result, increasing variance in b_0 would then *decrease* total community production.

A.4 Methods: Approximating the Shape of the Community Trait Distribution via Community-Weighted Measures

In order to assess predictions of TDT, it is necessary to quantify the full distribution of traits in a community, $C(z_i)$. This involves measuring the trait values of *all individuals* and thus incorporates both inter- and intraspecific trait variability. While measuring traits of *all* individuals in a community is ideal and several studies have done so (Albert et al., 2010; Gaucherand and Lavorel, 2007; Lavorel et al., 2008), it is a time-consuming work (Baraloto et al., 2010). While there are limitations, the trait biomass distribution, $C(z_i)$, can be approximated, and predictions of TDT can be tested without explicitly measuring the traits of all individuals.

Trait distributions can be approximated in two ways. The first method is straightforward and calculates the weighted trait distribution by taking the mean species trait value and multiplying by a measure of dominance (cover, biomass, abundance; Grime, 1998). This method can be implemented by calculating the central moments of joint distribution.

This community-weighted variance or CWM is increasingly a standard metric in trait-based ecology (Garnier and Navas, 2012; Lavorel, 2013; Violle et al., 2007) and represents the trait mean calculated for all species in a community weighted by species abundances as follows:

$$\text{CWM}_{j,\gamma} = \sum_{k=1}^{n_j} A_{k,j} \cdot z_k \quad (\text{A16})$$

where n_j is the number of species sampled in plot j , $A_{k,j}$ is the relative abundance of species k in plot j , and z_k is the mean value of species k . Several studies have also assessed the community-weighted variance of the trait distribution (see [Lavorel et al., 2011](#); [Ricotta and Moretti, 2011](#)).

The assemblage variance, V , calculated via the biomass-weighted values for the community-weighted variance ($CWV_{j,y}$) is given by

$$CWV_{j,y} = \sum_{k=1}^{n_j} A_{k,j} \cdot (z_k - CWM_{j,y})^2 \quad (\text{A17})$$

Further, the central moments skewness and kurtosis ($CWS_{j,y}$ and $CWK_{j,y}$, respectively) are given by

$$CWS_{j,y} = \frac{\sum_{k=1}^{n_j} A_{k,j} \cdot (z_k - CWM_{j,y})^3}{CWV_{j,y}^{3/2}};$$

$$CWK_{j,y} = \frac{\sum_{k=1}^{n_j} A_{k,j} \cdot (z_k - CWM_{j,y})^4}{CWV_{j,y}^2} - 3 \quad (\text{A18})$$

A limitation of this approach, however, is that it ignores the contribution of intraspecific trait variability. Community trait moments may also be sensitive to the distribution of abundances across species. For example, a highly positive community kurtosis value may just reflect the hyper-dominance of one species and not the true dispersion of traits again due to intraspecific variation.

A second method utilizes subsampling individuals to obtain a better approximation of how intraspecific variation influences the community distribution. By subsampling individuals for each species, one can begin to incorporate intraspecific variation around mean trait values for each species.

In [Fig. A5](#), we highlight a typical example that we believe can be used to generate two approximations of the community trait distribution. We use data from Konza Prairie LTER, Kansas, USA ([McAllister et al., 1998](#)). First, data were collected for the abundance of each species. These data are illustrated in [Fig. A4](#) to estimate the community trait distribution from mean and variance measure of species traits. We find that, consequently and counter-intuitively, the inclusion of intraspecific variation will likely simplify modeling efforts because these types of distributions are much easier to manipulate and understand analytically. For each species, the standard deviation of trait variation is equal to the reported standard error multiplied by the square root (where $n=3$). In sum, the community trait distribution can be

approximated in two ways (methods B and C). While B emphasizes interspecific variation, C also begins to include intraspecific variation. Method B is a reasonable approximation and can easily be implemented by most ecological studies as it only requires interspecific trait information and local abundance values. Method C requires an additional standardized subsampling method to estimate the standard error for each species but will result in a more accurate moment approximation.

A.5 Methods: Rocky Mountain Biological Lab: Shifts in Trait Distributions and Ecosystem Measures Across an Elevational Gradient

A.5.1 Measuring Whole-Community Trait Distributions

We measured community trait distributions and whole-ecosystem carbon flux data along an elevational gradient near Gothic, Colorado. The elevation gradient ranged between 2460 and 3380 m and spans 39 km in geographic distance. The elevational gradient contains five long-term study sites that run from dry, shrub-dominated high desert in Almont Colorado (2475 m) through the subalpine zone, to just below tree line (3380 m). The gradient consists of five long-term study sites that were established by Enquist in 2003 and has been sampled every year since. The gradient is located within Washington Gulch and East River valleys near the Rocky Mountain Biological Lab.

Each study site along the elevation gradient has similar local slope, aspect, and vegetation physiognomy. The sample area is approximately 50 m² and consists of a mixture of shrubs, grasses, and forbs. As discussed in [Bryant et al. \(2008\)](#), there is substantial turnover of plant species between sites with very few of the 120 species sampled, occurring in more than two of the sites. Additionally, shrub cover across the gradient decreases from a high of 33% at the lowest elevation site to 0% at the highest.

We utilized carbon flux data collected during the summer months of 2010 measured across the gradient. A species list and phylogeny for species at each site are given by [Bryant et al. \(2008\)](#). All sites contain weather stations on-site or nearby. Each study site has a similar local slope and south-southwest aspect and contains a mixture of herbaceous perennials, grasses, and shrubs. Since 2003, each year, five 1.3 × 1.3 m plots have been established haphazardly along the local slope of each study site, with at least 5 m distance between plots.

In 2010, Henderson measured the SLA of each plot and collected one fully expanded leaf from every individual. Fresh leaf samples were scanned

(with petiole) in the laboratory, then dried to a constant mass, and weighed. The trait values measured from every individual in each community were compiled to create individual-level trait distributions for SLA. In total, leaves from 2253 individuals across 54 species were collected and measured at the five sites. Species turnover was high, with only 11 species being found at more than one site and only one species found at more than two sites.

A.5.2 Gas Exchange and Productivity Measures

In 25 plots (5 plots per elevational site), we measured total ecosystem carbon flux. Carbon flux was measured as instantaneous daytime peak uptake (ca. 10 am) and night-time peak respiration (ca. 10 pm) (Saleska et al., 1999). Ambient CO₂ was measured by a Li-Cor 7500 infra red gas analyzer for 30 s and then the tent was put in place over the plot and the CO₂ concentration within the tent was measured for 90 s (Jasoni et al., 2005). Daytime measurements were only taken under cloudless conditions. The tent was designed to let in 75% of photosynthetically active radiation (tent fabric by Shelter Systems). Air inside the tent is well mixed by fans, and the tent chamber was sealed using a long skirt along the base of the tent that was covered with a heavy chain. The volume of the tent used along the gradient was 2.197 m³.

Soil efflux was measured at the same time as NEP using a Li-Cor 6400 portable photosynthesis machine with the soil chamber. The soil chamber fits inside a PVC soil collar, which was placed in the plot at least 2 weeks prior to the first measurement. Soil efflux was measured in two places in each plot along the gradient and one place per plot for the manipulation.

Carbon flux measurements along the elevational gradient were taken 4 weeks after snowmelt and then again at peak season (approximately 4 weeks after the first measurement, or when the majority of plants reached maximum height). Each NEP measurement consisted of daytime peak uptake (at ~10 am) and night-time respiration (at ~10 pm). Following the method of Jasoni et al. (2005), ambient CO₂ was measured for 30 s and then the tent was lowered and the CO₂ concentration within the tent was measured for 90 s, under clear sky and low wind conditions. Air inside the tent was well mixed by fans, and the chamber was sealed to the ground using a heavy chain.

For data analysis, we fit the predicted TDT model, using multiple regression in R using the 'car' library we fit the following linear model

$$\ln(\log_{10}(\text{NEP})) \sim \text{CWM.SLA} + \text{CWV.SLA} + \log_{10}(\text{Bio}) + \text{as.factor}(\text{Site})$$

where CWM.SLA is the community-weighted mean SLA and CWV.SLA is the community-weighted variance SLA calculated using the above equations for CWM and CWV as presented in the `vegan` package in R. Here, all values of SLA were log transformed before analyses, and $\log_{10}(\text{Bio})$ is the \log_{10} total above ground dry biomass at the time of carbon flux measurement. We use the site elevation of the sample as a factor in the model. The fit of this model explained a large fraction, $\sim 78\%$, of the variation in NEP ($R^2 = 0.778$, $df = 22$, $F = 11.04$, $p < 0.0001$, $AIC = -24.38$). To evaluate potential collinearity problems that may arise from linear relationships between model covariates (Ryan, 1997) we calculated variance-inflation factors (VIFs) for each covariate in each model using `vif()` from the package `car` in R. All VIFs were generally less than 5 except for one site where the $vif = 5.86$ which is nonetheless still low. In this model, the effect of CWV.SLA is significant ($p = 0.023$, $t = 2.45$, $SE = 0.072$, $parameter = 0.337$), but CWV.SLA and total biomass are marginally significant ($p = 0.068$, $t = -1.921$, $SE = 0.067$, $parameter = -0.138$; $p = 0.059$, $t = 1.994$, $SE = 0.067$, $parameter = 0.135$).

Variation in NEP across the gradient appears to be primarily due to the CWM and CWV of community SLA. Removing the parameter biomass and fitting a more simplified model with only mean and variance in SLA.

```
lm(log10(NEP) ~ CWM.SLA + CWV.SLA + as.factor(Site))
```

predicts a similar amount of variation in NEP to the full model above ($p < 0.0001$, Adjusted $R^2 = 0.67$, $AIC = -21.40$ and both CWM.SLA and CWV.SLA are now significant within the model ($p = 0.035$, $t = 2.247$, $SE = 0.358$, $parameter = 0.803$; $p = 0.0386$, $t = -2.194$, $SE = 0.075$, $parameter = -0.165$)).

Fitting a more simple model just using either plot biomass or plot CWM.SLA with site as a factor results in a poorer fit model when compared with the TDT-predicted model with lower R^2 and higher AIC values ($(\log_{10}(\text{posNEP}) \sim \log_{10}\text{Bio} + \text{as.factor}(\text{Site_name})$, $R^2 = 0.687$, $AIC = 17.994$; $\text{lm}(\log_{10}(\text{NEP}) \sim \text{CWM.SLA} + \text{as.factor}(\text{Site}))$; $R^2 = 0.684$, $AIC = -17.704$). Further, in both models, the effect of biomass and CWM.SLA was marginally significant ($p = 0.064$ and $p = 0.056$, respectively). These results indicate that together the CWM and CWV of community SLA are primary drivers of variation in community carbon flux.

A.6 Methods: PGE: Background, Methods, and Discussion

A.6.1 Background and Methods

The original purpose of PGE, started in 1856, was to investigate the effects of high levels of inorganic fertilizers and organic manure on hay production

relative to control treatments (see references within Crawley et al., 2005 for additional details on methodology). Our analyses mainly focused on the PGE trait dynamics of Plots 2 and 16. These plots were selected because of their contrasting botanical composition and species richness (Crawley et al., 2005; Harpole and Tilman, 2007). Plot 2 (became plot 2/2 in 1996) received farm yard manure between 1856 and 1863, but since then has received no further manure or fertilizer inputs, and is now considered to be a control plot. Plot 16, started in 1858, is a fertilized, unlimed plot that receives annual N, P, K, Na, and Mg applications (48 kg N ha⁻¹ as sodium nitrate in spring; mineral applied in winter: 35 kg P ha⁻¹ P as triple superphosphate, 225 kg K ha⁻¹ as potassium sulphate, 15 kg Na ha⁻¹ as sodium sulphate, and 10 kg Mg ha⁻¹ as magnesium sulphate). For the Park Grass dataset, we approximated the central moments of the community trait distribution, $C(z)$, for trait z within plot j and year y using Eqs. (A9)–(A11). We analyzed the time series of these plots in terms of changes in botanical composition, traits, and species richness. To focus on how experimentally paired *local* communities have responded over time, we highlighted plots 2 and 16 (the other plots also showed similar responses).

A.6.2 Assignment of Trait Values and Biomass-Weighted Trait Distributions

We assessed changes in SLA, seed size, and height. These traits have also been proposed to capture most functional and life history variation across species (Westoby, 1998; Westoby et al., 2002). Seed size is thought to characterize regenerative traits not associated with our trait-based growth model developed in Eq. (A7). Including a regeneration trait provides a basis to assess if other niche- or dispersal-based processes acting on other traits may be more important in structuring the community than traits associated with growth, dC/dt (see also discussion on effect and response traits; Suding et al., 2008). Further, variation in seed size should not directly influence our ecosystem-level predictions for dC_{Tot}/dt as this trait is not explicit in Eq. (A7). According to Eq. (A7), plant height (or size, C) can influence ecosystem NPP. So any large shifts in mean plant height would be important to note as well. Trait values are for populations sampled in United Kingdom. We used the first four central moments of $C(z)$ for plot j and year y to calculate the biomass-weighted mean, variance, skewness, and kurtosis.

Within this experiment, species abundance was measured by cutting aboveground biomass to ground level from six randomly located quadrats (50 × 25 cm) within each experimental and control plot. The plant material was then sorted into species, oven dried at 80 °C for 24 h, and the dry matter

determined (Crawley et al., 2005; Williams, 1978). For each plot, yields were estimated by weighing standing biomass (t/ha at 100% dry matter) from the whole plot, harvested in mid-June. The plots were originally cut by scythe, then by horse-drawn, and then tractor-drawn mowers (Williams, 1978; see these references for additional methodological detail of the PGE; Crawley et al., 2005).

Our analysis of additional fertilization and control plots at Park Grass (plots 3 and 14) also reveal similar differences in trait distributions (mean, variance, skew, and kurtosis) between the control and fertilized plots. In sum, for all of the Park Grass plots, our central conclusions do not change. We observed coordinated shifts in the functional trait distribution. In [Table A1](#), we show the correlations between the central moments of the trait distribution and species richness. These correlations include plots 2 (control) and 16 (fertilized) together. *None* of the central moments have significant correlation with species richness, indicating that the mechanisms and responses to environmental change captured by the shape of trait distributions are not captured by species richness. [Figure A1](#) shows the change in the central moments of the community trait distribution for SLA. [Figure A4](#) shows the associated changes in seed size in the 140-year long-term PGE. [Figure A5](#) shows the change in the central moments of the community trait distribution for adult height in the 140-year long-term PGE.

A.6.3 PGE—Additional Discussion

Fertilization also changed the shape of the SLA trait distribution indicating that the underlying forces that structure these communities under differing environments changed. For example, fertilization led to a reduction in the variance (the community mean SLA was negatively related to the

Table A1 Correlations Between the Central Moments of the Community Trait Distribution of Specific Leaf Area or SLA and Species Richness (Plots 2 (Control) and 16 (Fertilized) Together)

		Species Richness
SLA	Mean	0.00 ^{ns} .
	Variance	0.05 ^{ns} .
	Skewness	0.08 ^{ns} .
	Kurtosis	0.03 ^{ns} .

None of the moments have significant correlation with species richness, indicating that the mechanisms and responses to environmental change captured by the moments are not captured by species richness. This represents one of the great advantages for Trait Driver Theory (TDT) over theories based on species richness.

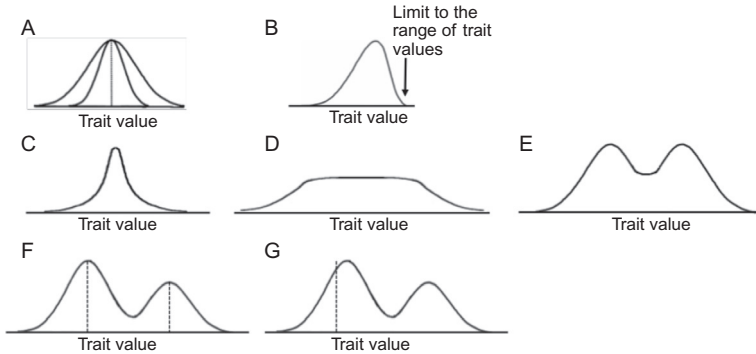


Figure A1 Trait Driver Theory and examples of the first four central moments of trait distributions: (A) *mean* (the first moment) and *variance* (the second moment). In this example, we show two communities with the same mean trait value (dashed line) but with different variances; (B) *skew* (a combination of the second and third moments). Skewness in a trait distribution can be caused by (i) time lags in community response to a new optimum trait value where a long tail of individuals expressing suboptimal trait values is present in the community (e.g. see Fig. 2), (ii) lopsided trait immigration into the community, or (iii) physical or physiological limits on trait expression (e.g. hydrological constraints on plant height); and (iv) may reflect rare species advantage. As shown in this example; and *Kurtosis* (a measure of the fourth moment relative to the second moment). Competitive exclusion and/or strong stabilizing selection will give a highly peaked (fourth moment kurtosis) distribution (C) while niche packing reflecting biotic interactions could give a more uniform distribution (D). *Note*: a normal trait distribution is defined by a skewness and kurtosis = 0. The more peaked the distribution, the more positive the kurtosis value (including the logistic, hyperbolic secant, and Laplace distributions). In contrast, processes that result in the ‘spreading out’ of traits will be characterized by increasingly more negative kurtosis values. In the case of a uniform distribution, kurtosis = -1.2. An increasingly bimodal distribution (Bernoulli distribution) will have kurtosis values = -2. (E) Bimodal distributions could arise where there are multiple optimal (dashed lines) trait values (F), or where the community is responding to a recent environmental change where the two peaks represent both an increased representation of nearly optimal individuals (the high peak) and the continued presence of individuals with optimal trait values for the historic environment (the low peak) (G). Dashed lines correspond to the optimal trait value(s).

community SLA variance, $r^2 = 0.48$, $p < 0.001$), suggesting either that fertilization was an environmental ‘filter’ on traits and/or competitive exclusion increased (see Table 2). The observed increase in the skewness and kurtosis of the SLA distribution with fertilization is in accord with predictions and expectations of TDT where more quick directional shifts in z_{opt} will lead to a skewed distribution. In contrast, the control plot trait distribution did not show dramatic changes in the variance or skewness of the distribution. However, the mean of SLA in the control plot did significantly

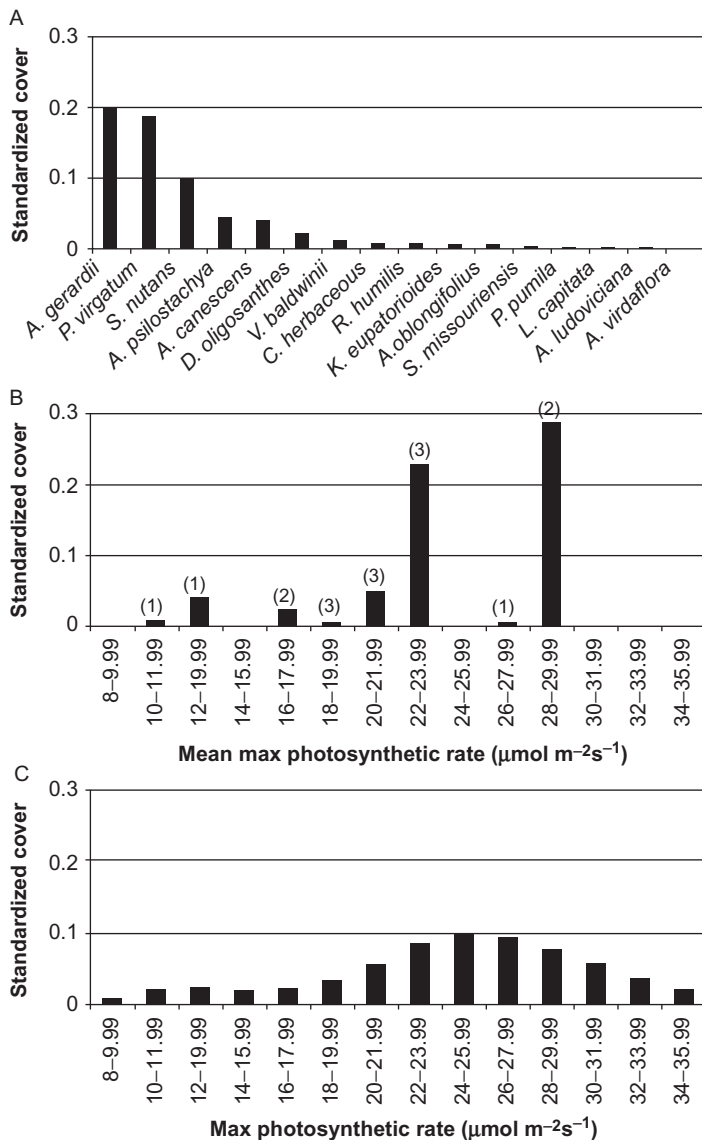


Figure A2 Examples of estimation of a community trait distribution from utilizing either mean trait values and/or intraspecific variation. In this example, we use abundance information (A) for the percent cover for 16 species from Konza Prairie LTER, Kansas, USA. Next, a trait measure, the leaf photosynthetic rate, was measured on a minimum of three leaves on three separate plants. Species means and standard errors were then calculated for this trait for each species. In (B) using the first method, the trait abundance distribution was calculated using only the mean trait data for the species in (A). Numbers in the parentheses indicate the number of species in each trait bin, and the peaks correspond to some of the dominant species in plot (A) that were rank ordered by abundance and not by photosynthetic rate. Lastly, using the second method, the community trait distribution of all individuals can be further approximated by integrating intraspecific subsampling. In (C), for each species, we incorporated intraspecific variation by using the standard error for this trait as measured in each species and then assumed a normal distribution around each species mean. We then generated the community-wide trait distribution by sampling from each species intraspecific trait distribution (defined by its mean and SE). The resulting distribution (C) is much more continuous and unimodal than in plot (B), which does not include intraspecific variation.

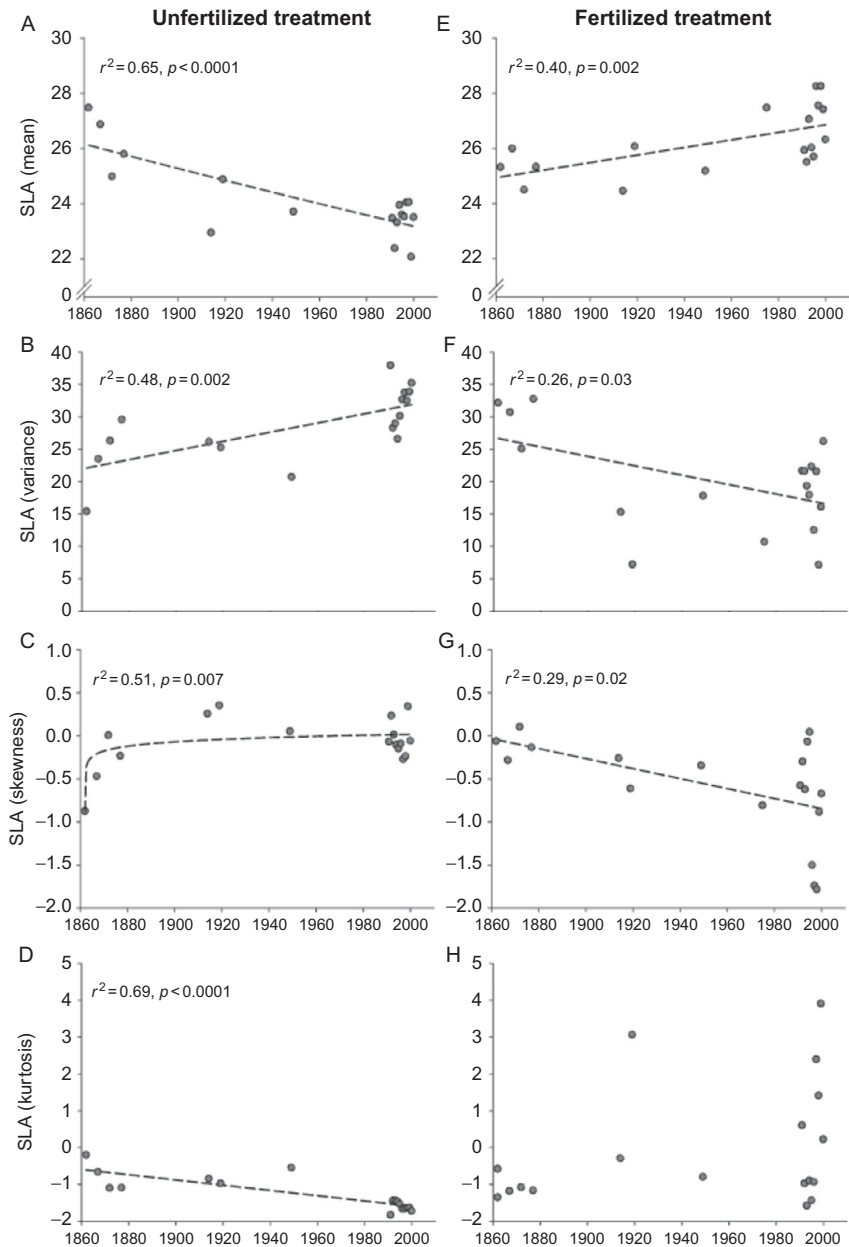


Figure A3 Change in the central moments of the community trait distribution for a key trait—specific leaf area or SLA—in the 140-year long-term Park Grass experiment. Regression lines are indicated for significant relationships. Fertilization has caused a decrease in the variance and an increase in the skewness. Fertilization increases the mean assemblage specific leaf area (SLA) but reduces the variance. This result indicates a directional shift in trait optimum, z_{opt} , and a functional shift in the composition of the community. Further, fertilized plots have become increasingly more skewed and have increasingly more positive kurtosis values indicating that communities have become increasingly dominated by a few trait values. Distributions with kurtosis values of -1.2 are characteristic of an ‘overdispersed’ uniform distribution, while plots with kurtosis values greater than 0 are more clumped/peaked than expected from a normal or Gaussian distribution.

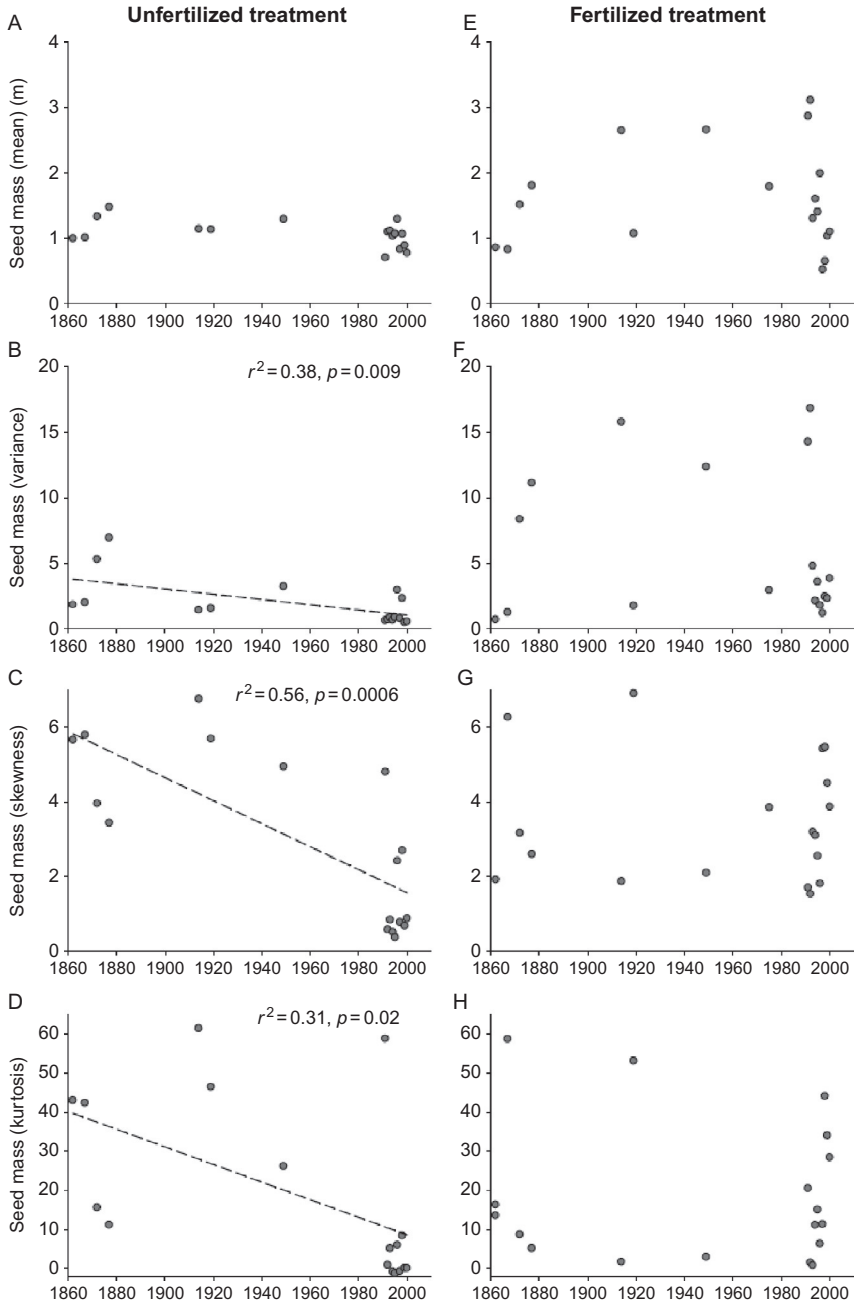


Figure A4 Change in the central moments of the community trait distribution for seed size in the 140-year long-term Park Grass experiment. Significant correlations are indicated with presence of fitted (dashed) regression lines.

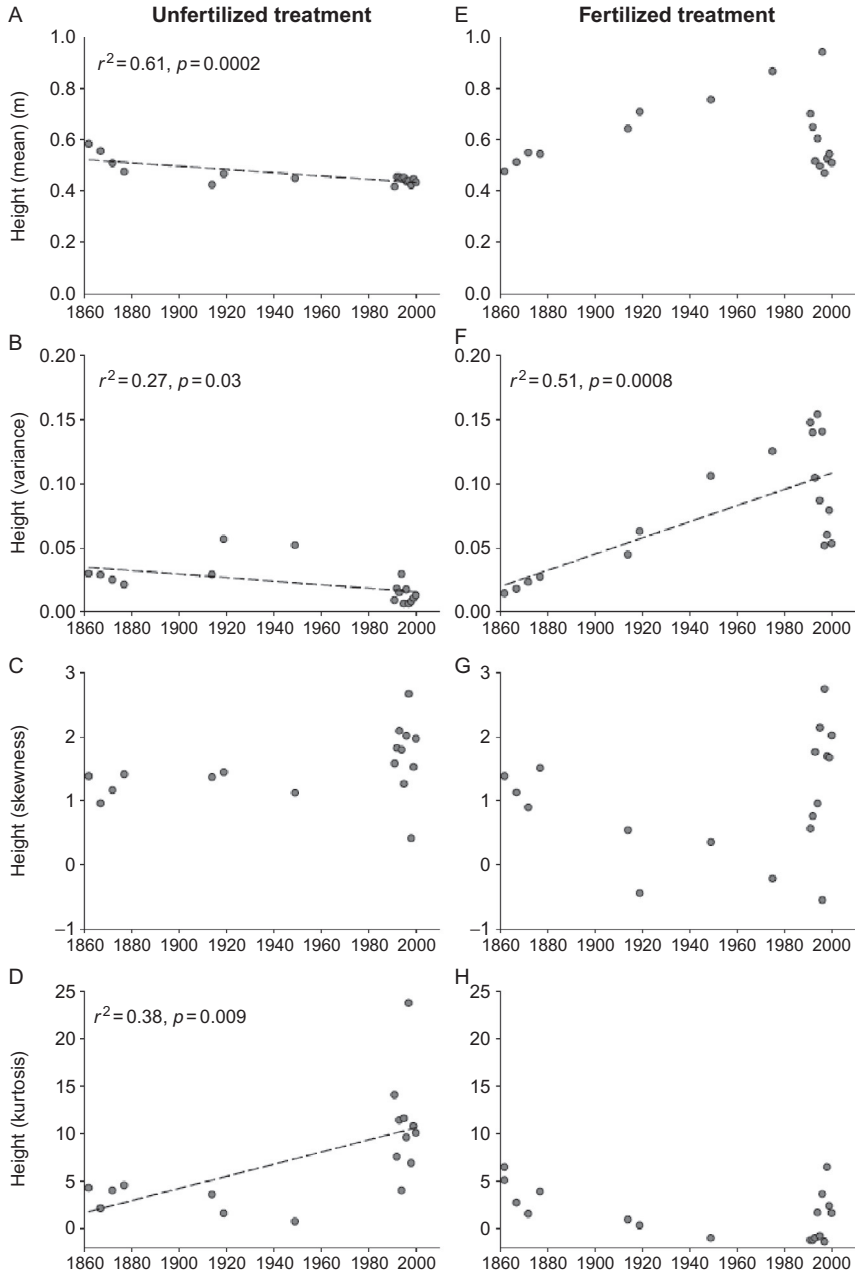


Figure A5 Change in the central moments of the community trait distribution for adult height in the 140-year long-term Park Grass experiment. Significant correlations are indicated with presence of fitted (dashed) regression lines.

decrease suggesting that natural and/or more gradual changes in the local environment (possibly due to an increase in nitrogen limitation over time and/or climate change) and/or recovery from past disturbance have influenced the control community. The kurtosis of the trait distribution in the control plot remained negative and close to -1.2 (a uniform distribution) consistent with an increased role of divergent ecological forces (niche packing and the role of biotic interactions). In contrast, in the fertilized plot, the variance of the distribution has decreased and the kurtosis tends to exhibit high positive values.

In the PGE, fertilization can be seen as a specific environmental driver, *E*. Fertilization changes soil resource availability and, according to TDT, differentially shifts the optimum growth rate. Indeed, in support we see a shift in z_{opt} (here being SLA) associated with fertilization. Analysis of the moments of distributions for two other community traits, seed size and adult reproductive height, shows that these trait means did not appreciably change with fertilization (Figs. A4 and A5). Importantly, the community mean of plant height did not change with fertilization supporting our assumption that observed change in community NPP was primarily due to changes in SLA, and also that the mean plant size or biomass, *C*, did not appreciably change. The one change with fertilization that we do observe is that the community variance of plant height increased. No other traits show any changes in the fertilized plots. Future work elaborating TDT should include the role of multiple trait drivers and their associated predictions.

Appendix references

- Abrams, P.A. & Chen, X. (2002) The effect of competition between prey species on the evolution of their vulnerabilities to a shared predator. *Evolutionary Ecology Research*, 897–909.
- Albert, C.H., Thuiller, W., Yoccoz, N., Soudan, A., Boucher, F., Saccone, P. & Lavorel, S. (2010) Intraspecific functional variability: extent, structure and sources of variation. *Journal of Ecology*, **98**, 604–613.
- Baraloto, C., Timothy Paine, C.E., Patiño, S., Bonal, D., Hérault, B. & Chave, J. Functional trait variation and sampling strategies in species-rich plant communities. *Functional Ecology*, **24**, 208–216.
- Brooker, R.W., Maestre, F.T., Callaway, R.M., Lortie, C.L., Cavieres, L.A., Kunstler, G., Liancourt, P., Tielbörger, K., Travis, J.M.J., Anthelme, F., Armas, C., Coll, L., Corcket, E., Delzon, S., Forey, E., Kikvidze, Z., Olofsson, J., Pugnaire, F., Quiroz, C.L., Saccone, P., Schifffers, K., Seifan, M., Touzard, B. & Michalet, R. (2008) Facilitation

- in plant communities: the past, the present, and the future. *Journal of Ecology*, **96**, 18–34.
- Chesson, P. (2000) Mechanisms of maintenance of species diversity. *Annual Review of Ecology and Systematics*, **31**, 343–366.
- Crawley, M.J., Johnston, A.E., Silvertown, J., Dodd, M., De Mazancourt, C., Heard, M.S., Henman, D.F. & Edwards, G.R. (2005) Determinants of species richness in the Park Grass Experiment. *American Naturalist*, **165**, 179–192.
- Diamond, J.M. (1975) Assembly of species communities. *Ecology and Evolution of Communities* (ed. by M.L. Cody and J.M. Diamond), pp. 342–444. Harvard Univ. Press, Cambridge, MA.
- Enquist, B.J., West, G.B. & Brown, J.H. (2009) Extensions and evaluations of a general quantitative theory of forest structure and dynamics. *Proceedings of the National Academy of Sciences of the United States of America*, **106**, 7046–7051.
- Freckleton, R.P. & Watkinson, A.R. (2001) Predicting competition coefficients for plant mixtures: reciprocity; transitivity and correlations with life-history traits. *Ecology Letters*, **4**, 348–357.
- Garnier, E. & Navas, M.-L. (2012) A trait-based approach to comparative functional plant ecology: concepts, methods and applications for agroecology. A review. *Agronomy for Sustainable Development*, **32**, 365–399.
- Gaucherand, S. & Lavorel, S. (2007) New method for rapid assessment of the functional composition of herbaceous plant communities. *Austral Ecology*, **32**, 927–936.
- Grime, J.P. (1998) Benefits of plant diversity to ecosystems: Immediate, filter and founder effects. *Journal of Ecology*, **86**, 902–910.
- Grime, J.P. (2006) Trait convergence and trait divergence in herbaceous plant communities: Mechanisms and consequences. *Journal of Vegetation Science*, **17**, 255–260.
- Harpole, W.S. & Tilman, D. (2007) Grassland species loss due to reduced niche dimension. *Nature*, **446**, 791–793.
- Hubbell, S.P. (2001) *The Unified Neutral Theory of Biodiversity and Biogeography*. Princeton University Press, Princeton, NJ.
- Karr, J.R. & James, F.C. (1975) Ecomorphological configurations and convergent evolution in species and communities. *Ecology and Evolution of Communities* (ed. by M.L. Cody and J.M. Diamond), pp. 258–291. Harvard University Press, Cambridge.
- Keddy, P.A. (1992) Assembly and response rules: two goals for predictive community ecology. *Journal of Vegetation Science*, **3**, 157–164.

- Kraft, N.J.B. & Ackerly, D.D. (2010) Functional trait and phylogenetic tests of community assembly across spatial scales in an Amazonian forest. *Ecological Monographs*, **80**, 401–422.
- Kraft, N.J.B., Valencia, R. & Ackerly, D.D. (2008) Functional traits and niche-based tree community assembly in an Amazonian forest. *Science*, **322**, 580–582.
- Kunstler, G., Albert, C.H., Courbaud, B., Lavergne, S., Thuiller, W., Vieilledent, G., Zimmermann, N.E. & Coomes, D.A. (2012) Effects of competition on tree radial-growth vary in importance but not in intensity along climatic gradients. *Journal of Ecology*, **99**, 300–312.
- Lavorel, S. (2013) Plant functional effects on ecosystem services. *Journal of Ecology*, **101** 4–8.
- Lavorel, S., Grigulis, K., Lamarque, P., Colace, M.-P., Garden, D., Girel, J., Pellet, G. & Douzet, R. (2011) Using plant functional traits to understand the landscape distribution of multiple ecosystem services. *Journal of Ecology*, **99**, 135–147.
- Lavorel, S., Grigulis, K., McIntyr, S., Williams, N., Garden, D., Dorr-ough, J., Berman, S., Qué-tier, F., Thebault, A. & Bonis, A. (2008) Assessing functional diversity in the field: methodology matters! *Functional Ecology*, **22**, 134–147.
- Levins, R. (1968) *Evolution in Changing Environments*. Princeton University Press, Princeton, N.J.
- MacArthur, R.H. (1958) Population ecology of some warblers of north-eastern coniferous forests. *Ecology*, **39**, 599–619.
- MacArthur, R.H. & Levins, R. (1967) The limiting similarity, convergence, and divergence of coexisting species. *The American Naturalist*, **101**, 377–385.
- Mayfield, M.M. & Levine, J.M. (2010) Opposing effects of competitive exclusion on the phylogenetic structure of communities. *Ecology Letters*, **13**, 1085–1093.
- McAllister, C.A., Knapp, A.K. & Maragni, L.A. (1998) Is leaf-level photosynthesis related to plant success in a highly productive grassland? *Oecologia*, **117**, 40–46.
- Mooney, H. (ed.) (1977) *Convergent evolution in Chile and California: Mediterranean climate ecosystems*. Dowden, Hutchinson and Ross, Stroudsburg, PA.
- Navas, M.-L. & Violle, C. (2009) Plant traits related to competition: how do they shape the functional diversity of communities? *Community Ecology*, **10**, 131–137.

- Orians, G.H. & Paine, R.T. (1983) Convergent evolution at the community level. *Coevolution* (ed. by D.J. Futuyma and M. Slatkin). Sinauer, Sunderland, MA.
- Poorter, H. (1989) Interspecific variation in relative growth rate: on ecological causes and physiological consequences. *Causes and Consequences of Variation in Growth Rate and Productivity in Higher Plants* (ed. by H. Lambers, M.L. Cambridge, H. Konings and T.L. Pons). SPB Academic Publishing, The Hague.
- Ricotta, C. & Moretti, M. (2011) CWM and Rao's quadratic diversity: a unified framework for functional ecology. *Oecologia*, **167**, 181-188.
- Savage, V.M. (2004) Improved approximations to scaling relationships for species, populations, and ecosystems across latitudinal and elevational gradients. *Journal of Theoretical Biology*, **227**, 525-534.
- Savage, V.M., Webb, C.T. & Norberg, J. (2007) A general multi-trait-based framework for studying the effects of biodiversity on ecosystem functioning. *Journal of Theoretical Biology*, **247**, 213-229.
- Schimper, A.F.W. (1898) *Pflanzengeographie auf Physiologischer Grundlage*, G. Fischer
- Schimper, A.F.W. (1903) *Plant-geography Upon a Physiological Basis*. Clarendon, Oxford, UK.
- Suding, K.N., Lavorel, S., Chapin, F.S., Cornelissen, J.H.C., Díaz, S., Garnier, E., Goldberg, D., Hooper, D.U., Jackson, S.T. & Navas, M.-L. (2008) Scaling environmental change through the community-level: a trait-based response-and-effect framework for plants. *Global Change Biology*, **14**, 1125-1140.
- Terborgh, J. (1973) On the notion of favorableness in plant ecology. *The American Naturalist*, **107**, 481-501.
- Tilman, D. (1982) *Resource Competition and Community Structure*. Princeton University Press, Princeton, NJ.
- Violle, C., Navas, M.-L., Vile, D., Kazakou, E., Fortunel, C., Hummel, I. & Garnier, E. (2007) Let the concept of trait be functional! *Oikos*, **116**, 882-892.
- Weiher, E. & Keddy, P.A. (1999) Assembly rules as general constraints on community composition. *Ecological Assembly Rules: Perspectives, Advances, Retreats* (ed. by E. Weiher, And Keddy, P.A.). Cambridge University Press, Cambridge, MASS.
- Westoby, M. (1998) A leaf-height-seed (LHS) plant ecology strategy scheme. *Plant and Soil*, **199**, 213-227.

- Westoby, M. & Wright, I.J. (2006) Land-plant ecology on the basis of functional traits. *Trends in Ecology & Evolution*, **21**, 261–268.
- Westoby, M., Falster, D.S., Moles, A.T., Vesk, P.A. & Wright, I.J. (2002) Plant ecological strategies: Some leading dimensions of variation between species. *Annual Review of Ecology and Systematics*, **33**, 125–159.
- Williams, E.D. (1978) *Botanical Composition of the Park Grass plots at Rothamsted 1856-1976*. Rothamsted Experimental Station, Harpenden, United Kingdom.

REFERENCES

- Ackerly, D.D., 2003. Community assembly, niche conservatism, and adaptive evolution in changing environments. *Int. J. Plant Sci.* 164, S165–S184.
- Ackerly, D.D., Monson, R.K., 2003. Waking the sleeping giant: the evolutionary foundations of plant function. *Int. J. Plant Sci.* 164, S1–S6.
- Adler, P.B., Salguero-Gómez, R., Compagnoni, A., Hsu, J.S., Ray-Mukherjee, J., Mbeau-Ache, C., Franco, M., 2014. Functional traits explain variation in plant life history strategies. *Proc. Natl. Acad. Sci. U.S.A.* 111, 740–745.
- Albert, C.H., Thuiller, W., Yoccoz, N., Soudan, A., Boucher, F., Saccone, P., Lavorel, S., 2010. Intraspecific functional variability: extent, structure and sources of variation. *J. Ecol.* 98, 604–613.
- Antonovics, J., 1976. The nature of limits to natural selection. *Ann. Mo. Bot. Gard.* 63, 224–227.
- Arnold, S.J., 1983. Morphology, performance, and fitness. *Am. Zool.* 23, 347–361.
- Asner, G.P., Martin, R.E., Tupayachi, R., Anderson, C.B., Sinca, F., Carranza-Jiménez, L., Martínez, P., 2014. Amazonian functional diversity from forest canopy chemical assembly. *Proc. Natl. Acad. Sci. U.S.A.* 111, 5604–5609.
- Baraloto, C., Timothy Paine, C.E., Patiño, S., Bonal, D., Hérault, B., Chave, J., 2010. Functional trait variation and sampling strategies in species-rich plant communities. *Funct. Ecol.* 24, 208–216.
- Belmaker, J., Jetz, W., 2013. Spatial scaling of functional structure in bird and mammal assemblages. *Am. Nat.* 181, 464–478.
- Boulangéat, I., Philippe, P., Abdulhak, S., Douzet, R., Garraud, L., Lavergne, S., Lavorel, S., Van Es, J., Vittoz, P., Thuiller, W., 2012. Improving plant functional groups for dynamic models of biodiversity: at the crossroads between functional and community ecology. *Glob. Chang. Biol.* 18, 3464–3475.
- Bryant, J.A., Lamanna, C., Morlon, H., Kerkhoff, A.J., Enquist, B.J., Green, J.L., 2008. Microbes on mountainsides: contrasting elevational patterns of bacterial and plant diversity. *Proc. Natl. Acad. Sci. U.S.A.* 105, 11505–11511.
- Cardinale, B.J., Wright, J.P., Cadotte, M.W., Carroll, I.T., Hector, A., Srivastava, D.S., Loreau, M., Weis, J.J., 2007. Impacts of plant diversity on biomass production increase through time because of species complementarity. *Proc. Natl. Acad. Sci. U.S.A.* 104, 18123–18128.
- Cavender-Bares, J., Kozak, K., Fine, P., Kembel, S., 2009. The merging of community ecology and phylogenetic biology. *Ecol. Lett.* 12, 693–715.
- Chapin, F.S.I., 1980. The mineral nutrition of wild plants. *Annu. Rev. Ecol. Syst.* 11, 233–260.

- Chapin, F.S., Shaver, G.R., 1985. Individualistic growth response of tundra plant species to environmental manipulations in the field. *Ecology* 66, 564–576.
- Chapin, F.S.I., Zavaleta, E.S., Eviner, V.T., Naylor, R.L., Vitousek, P.M., Reynolds, H.L., Hooper, D.U., Lavorel, S., Sala, O.E., Hobbie, S.E., Mack, M.C., Diaz, S., 2000. Consequences of changing biodiversity. *Nature* 405, 234–242.
- Choler, P., 2005. Consistent shifts in alpine plant traits along a mesotopographical gradient. *Arct. Antarct. Alp. Res.* 37, 444–453.
- Coomes, D.A., 2006. Challenges to the generality of WBE theory. *Trends Ecol. Evol.* 21, 593–596.
- Cornwell, W., Ackerly, D.D., 2009. Community assembly and shifts in plant trait distributions across an environmental gradient in coastal California. *Ecol. Monogr.* 79, 109–126.
- Craine, J.M., 2009. *Resource Strategies of Wild Plants*. Princeton University Press, Princeton.
- Craine, J.M., Froehle, J., Tilman, D.G., Wedin, D.A., Chapin, I.F.S., 2001. The relationships among root and leaf traits of 76 grassland species and relative abundance along fertility and disturbance gradients. *Oikos* 93, 274–285.
- Damuth, J., 1981. Population-density and body size in mammals. *Nature* 290, 699–700.
- Davis, M.B., Shaw, R.G., 2001. Range shifts and adaptive responses to quaternary climate change. *Science* 292, 673–679.
- Dell, A.I., Pawar, S., Savage, V.M., 2013. The thermal dependence of biological traits. *Ecology* 94, 1205–1206.
- Dell, A.I., Zhao, L., Brose, U., Pearson, R.G., Alford, R.A., 2015. Population and community body size structure across a complex environmental gradient. *Adv. Ecol. Res.* 52, 115–167.
- DeLucia, E., Drake, J.E., Thomas, R.B., Gonzalez-Meler, M., 2007. Forest carbon use efficiency: is respiration a constant fraction of gross primary production? *Glob. Chang. Biol.* 13, 1157–1167.
- Diaz, S., Cabido, M., 2001. Vive la difference; plant functional diversity matters to ecosystem processes. *Trends Ecol. Evol.* 16, 646–655.
- Diaz, S., Hodgson, J.G., Thompson, K., Cabido, M., Cornelissen, J.H.C., Jalili, A., Monserrat-Marti, G., Grime, J.P., Zarrinkamar, F., Asri, S., Band, R., Basconcelo, S., Castro-Diez, P., Funes, G., Hamzehee, B., Khoshnevi, M., Perez-Harguindeguy, N., Perez-Rontome, M.C., Shirvany, F.A., Vendramini, F., Yazdani, S., Abbas-Azimi, R., Bogaard, A., Boustani, S., Charles, M., Dehghan, M., de Torres-Espuny, L., Falczuk, V., Guerrero-Campo, J., Hynd, A., Jones, G., Kowsary, E., Kazemi-Saeed, F., Maestro-Martinez, M., Romo-Diez, A., Shaw, S., Siavash, B., Villar-Salvador, P., Zak, M.R., 2004. The plant traits that drive ecosystems: evidence from three continents. *J. Veg. Sci.* 15, 295–304.
- Diaz, S., Lavorel, S., De Bello, F., Quétier, F., Grigulis, K., Robson, M., 2007. Incorporating plant functional diversity effects in ecosystem service assessments. *Proc. Natl. Acad. Sci. U.S.A.* 36, 20684–20689.
- Dieckmann, U., Ferrière, R., 2004. Adaptive dynamics and evolving biodiversity. In: Ferrière, R., Dieckmann, U., Couvet, D. (Eds.), *Evolutionary Conservation Biology*. Cambridge University Press, Cambridge, UK, pp. 188–224.
- Doughty, C.E., Asner, G.P., Martin, R.E., 2011. Predicting tropical plant physiology from leaf and canopy spectroscopy. *Oecologia* 165, 289–299.
- Elser, J.J., Fagan, W.F., Kerkhoff, A.J., Swenson, N.G., Enquist, B.J., 2010. Biological stoichiometry of plant production: metabolism, scaling and ecological response to global change. *New Phytol.* 186, 593–608.
- Enquist, B.J., 2010. Wanted: a general and predictive theory for trait-based plant ecology. *Bioscience* 60, 854–855.

- Enquist, B.J., Brown, J.H., West, G.B., 1998. Allometric scaling of plant energetics and population density. *Nature* 395, 163.
- Enquist, B.J., Economo, E.P., Huxman, T.E., Allen, A.P., Ignace, D.D., Gillooly, J.F., 2003. Scaling metabolism from organisms to ecosystems. *Nature* 423, 639–642.
- Enquist, B.J., Kerkhoff, A.J., Huxman, T.E., Economo, E.P., 2007a. Adaptive differences in plant physiology and ecosystem invariants: insights from a metabolic scaling model. *Glob. Chang. Biol.* 13, 591–609.
- Enquist, B.J., Kerkhoff, A.J., Stark, S.C., Swenson, N.G., McCarthy, M.C., Price, C.A., 2007b. A general integrative model for scaling plant growth, carbon flux, and functional trait spectra. *Nature* 449, 218–222.
- Enquist, B.J., Allen, A.P., Brown, J.H., Gillooly, J.F., Kerkhoff, A.J., Niklas, K.J., Price, C.A., West, G.B., 2007c. Biological scaling: does the exception prove the rule? *Nature* 445, E9–E10.
- Enquist, B.J., West, G.B., Brown, J.H., 2009. Extensions and evaluations of a general quantitative theory of forest structure and dynamics. *Proc. Natl. Acad. Sci. U.S.A.* 106, 7046–7051.
- Evans, G.C., 1972. *The Quantitative Analysis of Plant Growth*. University of California Press, Berkeley.
- Ferriere, R., Legendre, S., 2013. Eco–evolutionary feedbacks, adaptive dynamics and evolutionary rescue theory. *Philos. Trans. R. Soc. B Biol. Sci.* 368, 20120081.
- Fonseca, C.R., Overton, J.M., Collins, B., Westoby, M., 2000. Shifts in trait–combinations along rainfall and phosphorus gradients. *J. Ecol.* 88, 964–977.
- Freckleton, R.P., Watkinson, A.R., 2001. Predicting competition coefficients for plant mixtures: reciprocity; transitivity and correlations with life–history traits. *Ecol. Lett.* 4, 348–357.
- Frenne, P., Graae, B.J., Rodríguez–Sánchez, F., Kolb, A., Chabrierie, O., Decocq, G., Kort, H., Schrijver, A., Diekmann, M., Eriksson, O., 2013. Latitudinal gradients as natural laboratories to infer species’ responses to temperature. *J. Ecol.* 101 (3), 784–795.
- Funk, J.L., Cleland, E.E., Suding, K.N., Zavaleta, E.S., 2008. Restoration through reassembly: plant traits and invasion resistance. *Trends Ecol. Evol.* 23, 695–703.
- Garnier, E., Navas, M.–L., 2012. A trait–based approach to comparative functional plant ecology: concepts, methods and applications for agroecology. A review. *Agron. Sustain. Dev.* 32, 365–399.
- Garnier, E., Cortez, J., Billes, G., Navas, M.–L., Roumet, C., Debussche, M., Laurent, G., Blanchard, A., Aubry, D., Bellmann, A., Neill, C., Toussaint, J.–P., 2004. Plant functional markers capture ecosystem properties during secondary succession. *Ecology* 85, 2630–2637.
- Gaucherand, S., Lavorel, S., 2007. New method for rapid assessment of the functional composition of herbaceous plant communities. *Austral Ecol.* 32, 927–936.
- Ghalambor, C.K., McKay, J.K., Carroll, S.P., Reznick, D.N., 2007. Adaptive versus non–adaptive phenotypic plasticity and the potential for contemporary adaptation in new environments. *Funct. Ecol.* 21, 394–407.
- Gilbert, J.P., DeLong, J.P., 2015. Individual variation decreases interference competition but increases species persistence. *Adv. Ecol. Res.* 52, 45–64.
- Gillooly, J.F., Brown, J.H., West, G.B., Savage, V.M., Charnov, E.L., 2001. Effects of size and temperature on metabolic rate. *Science* 293, 2248–2251.
- Gillooly, J., Allen, A.P., West, G.B., Brown, J.H., 2005. The rate of DNA evolution: effects of body size and temperature on the molecular clock. *Proc. Natl. Acad. Sci. U.S.A.* 102, 140–145.
- Goldberg, D.E., Landa, K., 1991. Competitive effect and response: hierarchies and correlated traits in the early stages of competition. *J. Ecol.* 79, 1013–1030.

- Grace, J.B., Michael Anderson, T., Smith, M.D., Seabloom, E., Anelman, S.J., Meche, G., Weiher, E., Allain, L.K., Jutila, H., Sankaran, M., 2007. Does species diversity limit productivity in natural grassland communities? *Ecol. Lett.* 10, 680–689.
- Grime, J.P., 1977. Evidence for the existence of three primary strategies in plants and its relevance to ecological and evolutionary theory. *Am. Nat.* 111, 1169–1194.
- Grime, J.P., 1998. Benefits of plant diversity to ecosystems: immediate, filter and founder effects. *J. Ecol.* 86, 902–910.
- Grime, J.P., 2006. Trait convergence and trait divergence in herbaceous plant communities: mechanisms and consequences. *J. Veg. Sci.* 17, 255–260.
- Grime, J.P., Hunt, R., 1975. Relative growth-rate: its range and adaptive significance in a local flora. *J. Ecol.* 63, 393–422.
- Han, W., Fang, J., Guo, D., 2005. Leaf N and P stoichiometry across 753 terrestrial plant species in China. *New Phytol.* 168, 377–385.
- Hillebrand, H., Matthiessen, B., 2009. Biodiversity in a complex world: consolidation and progress in functional biodiversity research. *Ecol. Lett.* 12, 1405–1419.
- Hooper, D.U., Chapin, F.S.I., Ewel, J.J., Hector, A., Inchausti, P., Lavorel, S., Lawton, J.H., Lodge, D.M., Loreau, M., Naeem, S., Schmid, B., Setälä, H., Symstad, A.J., Vandermeer, J., Wardle, D.A., 2004. Effects of biodiversity on ecosystem functioning: a consensus of current knowledge. *Ecol. Monogr.* 75, 3–35.
- Hubbell, S.P., 2001. *The Unified Neutral Theory of Biodiversity and Biogeography*. Princeton University Press, Princeton, NJ.
- Hulshof, C.M., Violle, C., Spasojevic, M.J., McGill, B., Damschen, E., Harrison, S., Enquist, B.J., 2013. Intra-specific and inter-specific variation in specific leaf area reveal the importance of abiotic and biotic drivers of species diversity across elevation and latitude. *J. Veg. Sci.* 24, 921–931.
- Jasoni, R.L., Smith, S.D., Arnone, J.A., 2005. Net ecosystem CO₂ exchange in Mojave Desert shrublands during the eighth year of exposure to elevated CO₂. *Glob. Chang. Biol.* 11, 749–756.
- Jung, V., Violle, C., Mondy, C., Hoffmann, L., Muller, S., 2010. Intraspecific variability and trait-based community assembly. *J. Ecol.* 98, 1134–1140.
- Kattge, J., Díaz, S., Lavorel, S., Prentice, I.C., Leadley, P., Bönsch, G., Garnier, E., Westoby, M., Reich, P.B., Wright, I.J., Cornelissen, J.H.C., Violle, C., Harrison, S.P., van Bodegom, P.M., Reichstein, M., Enquist, B.J., Soudzilovskaia, N.A., Ackerly, D.-D., Anand, M., Atkin, O., Bahn, M., Baker, T.R., Baldocchi, D., Bekker, R., Blanco, C.C., Blonder, B., Bond, W.J., Bradstock, R., Bunker, D.E., Casanoves, F., Cavender-Bares, J., Chambers, J.Q., Chapin III, F.S., Chave, J., Coomes, D., Cornwell, W.K., Craine, J.M., Dobrin, B.H., Duarte, L., Durka, W., Elser, J., Esser, G., Estiarte, M., Fagan, W.F., Fang, J., Fernández-Méndez, F., Fidelis, A., Finegan, B., Flores, O., Ford, H., Frank, D., Freschet, G.T., Fyllas, N.M., Gallagher, R.V., Green, W.A., Gutierrez, A.G., Hickler, T., Higgins, S.I., Hodgson, J.-G., Jalili, A., Jansen, S., Joly, C.A., Kerkhoff, A.J., Kirkup, D., Kitajima, K., Kleyer, M., Klotz, S., Knops, J.M.H., Kramer, K., Kühn, I., Kurokawa, H., Laughlin, D., Lee, T.D., Leishman, M., Lens, F., Lenz, T., Lewis, S.L., Lloyd, J., Lusà, J., Louault, F., Ma, S., Mahecha, M.D., Manning, P., Massad, T., Medlyn, B.E., Messier, J., Moles, A.T., Müller, S.C., Nadrowski, K., Naeem, S., Niinemets, Ü., Nöllert, S., Nüske, A., Ogaya, R., Oleksyn, J., Onipchenko, V.G., Onoda, Y., Ordoñez, J., Overbeck, G., Ozinga, W.A., Patiño, S., Paula, S., Pausas, J.G., Peñuelas, J., Phillips, O.L., Pillar, V., Poorter, H., Poorter, L., Poschlod, P., Prinzing, A., Proulx, R., Rammig, A., Reinsch, S., Reu, B., Sack, L., Salgado-Negret, B., Sardans, J., Shiøder, S., Shipley, B., Siefert, A., Sosinski, E., Soussana, J.F., Swaine, E., Swenson, N., Thompson, K., Thornton, P., Waldram, M., Weiher, E.,

- White, M., White, S., Wright, S.J., Yguel, B., Zaehle, S., Zanne, A.E., Wirth, C., 2011. TRY—a global database of plant traits. *Glob. Chang. Biol.* 17, 2905–2935.
- Kingsolver, J.G., Huey, R.B., 2003. Introduction: The evolution of morphology, performance, and fitness. *Integr. Comp. Biol.* 43, 361–366.
- Kleyer, M., Bekker, R.M., Knevel, I.C., Bakker, J.P., Thompson, K., Sonnenschein, M., Poschod, P., van Groenendael, J.M., Klimes, L., Klimesova, J., Klotz, S., Rusch, G.M., Hermy, M., Adriaens, D., Boedeltje, G., Bossuyt, B., Dannemann, A., Endels, P., Gotzenberger, L., Hodgson, J.G., Jackel, A.K., Kuhn, I., Kunzmann, D., Ozinga, W.A., Romermann, C., Stadler, M., Schlegelmilch, J., Steendam, H.J., Tackenberg, O., Wilmann, B., Cornelissen, J.H.C., Eriksson, O., Garnier, E., Peco, B., 2008. The LEDA Traitbase: a database of life-history traits of Northwest European flora. *J. Ecol.* 96, 1266–1274.
- Knops, J.M., Reinhart, K., 2000. Specific leaf area along a nitrogen fertilization gradient. *Am. Midl. Nat.* 144, 265–272.
- Kraft, N.J.B., Valencia, R., Ackerly, D.D., 2008. Functional traits and niche-based tree community assembly in an Amazonian forest. *Science* 322, 580–582.
- Kunstler, G., Albert, C.H., Courbaud, B., Lavergne, S., Thuiller, W., Vieilledent, G., Zimmermann, N.E., Coomes, D.A., 2012. Effects of competition on tree radial-growth vary in importance but not in intensity along climatic gradients. *J. Ecol.* 99, 300–312.
- Lamanna, C.A., Blonder, B., Violle, C., Kraft, N.J.B., Sandel, B., Simova, I., Donoghue II, J.C., Svenning, J.-C., McGill, B.J., Boyle, B., Dolins, S., Jørgensen, P.M., Marcuse-Kubitza, A., Morueta-Holme, N., Peet, R.K., Piel, W., Regetz, J., Schildhauer, M., Spencer, N., Theirs, B.M., Wiser, S.K., Enquist, B.J., 2014. Functional trait space and the latitudinal diversity gradient. *Proc. Natl. Acad. Sci. U.S.A.* 111 (38), 13745–13750.
- Lambers, H., Freijns, N., Poorter, H., Hirose, T., van der Werff, H., 1989. Analyses of growth based on net assimilation rate and nitrogen productivity: their physiological background. In: Lambers, H., Cambridge, M.L., Konings, H., Pons, T.L. (Eds.), *Variation in Growth Rate and Productivity of Higher Plants*. SPB Academic Publishing, The Hague, The Netherlands, pp. 1–17.
- Laskowski, K.L., Pearish, S., Bensky, M., Bell, A.M., 2015. Predictors of individual variation in movement in a natural population of threespine stickleback (*Gasterosteus aculeatus*). *Adv. Ecol. Res.* 52, 65–90.
- Lavorel, S., Garnier, E., 2002. Predicting changes in community composition and ecosystem functioning from plant traits: revisiting the Holy Grail. *Funct. Ecol.* 16, 545–556.
- Lavorel, S., Díaz, S., Cornelissen, J.H.C., Garnier, E., Harrison, S.P., McIntyre, S., Pausas, J.G., Pérez-Harguindeguy, N., Roumet, C., Urcelay, C., 2007. Plant functional types: are we getting any closer to the Holy Grail? In: *Terrestrial Ecosystems in a Changing World*. Springer, Berlin Heidelberg, pp. 149–164.
- Lavorel, S., Grigulis, K., McIntyre, S., Williams, N., Garden, D., Dorrough, J., Berman, S., Quétiér, F., Thebault, A., Bonis, A., 2008. Assessing functional diversity in the field: methodology matters!. *Funct. Ecol.* 22, 134–147.
- Lavorel, S., Grigulis, K., Lamarque, P., Colace, M.-P., Garden, D., Girel, J., Pellet, G., Douzet, R., 2011. Using plant functional traits to understand the landscape distribution of multiple ecosystem services. *J. Ecol.* 99, 135–147.
- Levins, R., 1968. *Evolution in Changing Environments*. Princeton University Press, Princeton, NJ.
- Litchman, E., Klausmeier, C.A., 2008. Trait-based community ecology of phytoplankton. *Annu. Rev. Ecol. Evol. Syst.* 39, 615–639.
- Loreau, M., Naeem, S., Inchausti, P., Bengtsson, J., Grime, J.P., Hector, A., Hooper, D.U., Huston, M.A., Raffaelli, D., Schmid, B., Tilman, D., Wardle, D.A., 2001. Biodiversity and ecosystem functioning: current knowledge and future challenges. *Science* 294, 804–808.

- MacArthur, R.H., 1972. *Geographical Ecology: Patterns in the Distribution of Species*. Princeton University Press, Princeton, NJ.
- Marks, C., Lechowicz, M., 2006. Alternative designs and the evolution of functional diversity. *Am. Nat.* 167, 55–67.
- Mason, N.W.H., Moullot, D., Lee, W.G., Wilson, J.B., 2005. Functional richness, functional evenness and functional divergence: the primary components of functional diversity. *Oikos* 111, 112–118.
- Mayfield, M.M., Levine, J.M., 2010. Opposing effects of competitive exclusion on the phylogenetic structure of communities. *Ecol. Lett.* 13, 1085–1093.
- Maynard Smith, J., 1976. What determines the rate of evolution? *Am. Nat.* 110, 331–338.
- McGill, B.J., Enquist, B.J., Weiher, E., Westoby, M., 2006. Rebuilding community ecology from functional traits. *Trends Ecol. Evol.* 21, 178–185.
- Messier, J., McGill, B.J., Lechowicz, M.J., 2010. How do traits vary across ecological scales? A case for trait-based ecology. *Ecol. Lett.* 13, 838–848.
- Michaletz, S.T., Cheng, D., Kerkhoff, A.J., Enquist, B.J., 2014. Convergence of terrestrial plant production across global climate gradients. *Nature* 512, 39–43.
- Moses, M.E., Hou, C., Woodruff, W.H., West, G.B., Nekola, J.C., Zuo, W., Brown, J.H., 2008. Revisiting a model of ontogenetic growth: estimating model parameters from theory and data. *Am. Nat.* 171, 632–645.
- Muller, E.B., Nisbet, R.M., Kooijman, S., Elser, J.J., McCauley, E., 2001. Stoichiometric food quality and herbivore dynamics. *Ecol. Lett.* 4, 519–529.
- Naeem, S., Wright, J.P., 2003. Disentangling biodiversity effects on ecosystem functioning: deriving solutions to a seemingly insurmountable problem. *Ecol. Lett.* 6, 567–579.
- Naeem, S., Thompson, L.J., Lawler, S.P., Lawton, J.H., Woodfin, R.M., 1994. Declining biodiversity can alter the performance of ecosystems. *Nature* 368, 734–737.
- Naeem, S., Bunker, D.E., Hector, A., Loreau, M., Perrings, C., 2009. Can we predict the effects of global change on biodiversity loss and ecosystem functioning? In: Naeem, S., Bunker, D.E., Hector, A., Loreau, M., Perrings, C. (Eds.), *Biodiversity, Ecosystem Functioning, and Human Wellbeing: An Ecological and Economic Perspective*. Oxford University Press, Oxford, pp. 290–298.
- Norberg, J., 2004. Biodiversity and ecosystem functioning: a complex adaptive systems approach. *Limnol. Oceanogr.* 49, 1269–1277.
- Norberg, J., Swaney, D.P., Dushoff, J., Lin, J., Casagrandi, R., Levin, S.A., 2001. Phenotypic diversity and ecosystem functioning in changing environments: a theoretical framework. *Proc. Natl. Acad. Sci. U.S.A.* 98, 11376–11381.
- Paine, C., Baraloto, C., Chave, J., Hérault, B., 2011. Functional traits of individual trees reveal ecological constraints on community assembly in tropical rain forests. *Oikos* 120, 720–727.
- Pawar, S., 2015. The role of body size variation in community assembly. *Adv. Ecol. Res.* 52, 201–248.
- Petchey, O.L., Gaston, K.J., 2002. Functional diversity (FD), species richness and community composition. *Ecol. Lett.* 5, 402–411.
- Peters, R.H., 1983. *The Ecological Implications of Body Size*. Cambridge University Press, Cambridge.
- Pettorelli, N., Hilborn, A., Duncan, C., Durant, S.M., 2015. Individual variability: the missing component to our understanding of predator-prey interactions. *Adv. Ecol. Res.* 52, 19–44.
- Poorter, H., 1989. Interspecific variation in relative growth rate: on ecological causes and physiological consequences. *Causes and consequences of variation in growth rate and productivity of higher plants*, 45–68.
- Poorter, H., Niinemets, Ü., Poorter, L., Wright, I.J., Villar, R., 2009. Causes and consequences of variation in leaf mass per area (LMA): a meta-analysis. *New Phytol.* 182 (3), 565–588.

- Pulliam, R.H., 1988. Sources, sinks, and population regulation. *Am. Nat.* 132, 652–661.
- Reich, P.B., 2005. Global biogeography of plant chemistry: filling in the blanks. *New Phytol.* 168, 263–266.
- Reich, P.B., 2014. The world-wide ‘fast–slow’ plant economics spectrum: a traits manifesto. *J. Ecol.* 102, 275–301.
- Reich, P.B., Oleksyn, J., 2004. Global patterns of plant leaf N and P in relation to temperature and latitude. *Proc. Natl. Acad. Sci. U.S.A.* 101, 11001–11006.
- Reich, P.B., Walters, M.B., Ellsworth, D.S., 1997. From tropics to tundra: global convergence in plant functioning. *Proc. Natl. Acad. Sci. U.S.A.* 94, 13730–13734.
- Ricker, W.E., 1979. Growth rates and models. In: *Fish Physiology*. Academic Press, New York, pp. 677–743.
- Roscher, C., Schumacher, J., Gubsch, M., Lipowsky, A., Weigelt, A., Buchmann, N., Schmid, B., Schulze, E.-D., 2012. Using plant functional traits to explain diversity–productivity relationships. *PLoS One* 7, e36760.
- Rosenzweig, M.L., 1995. *Species Diversity in Space and Time*. Cambridge University Press, Cambridge.
- Ryan, T.P., 1997. *Modern Regression Methods*. Wiley.
- Safi, K., Cianciaruso, M.V., Loyola, R.D., Brito, D., Armour-Marshall, K., Diniz-Filho, J.A.F., 2011. Understanding global patterns of mammalian functional and phylogenetic diversity. *Philos. Trans. R. Soc. B Biol. Sci.* 366, 2536–2544.
- Saleska, S.R., Harte, J., Torn, M.S., 1999. The effect of experimental ecosystem warming on CO₂ fluxes in a montane meadow. *Glob. Chang. Biol.* 5, 125–141.
- Savage, V.M., 2004. Improved approximations to scaling relationships for species, populations, and ecosystems across latitudinal and elevational gradients. *J. Theor. Biol.* 227, 525–534.
- Savage, V.M., Gillooly, J.F., Brown, J.H., West, G.B., Charnov, E.L., 2004. Effects of body size and temperature on population growth. *Am. Nat.* 163, E429–E441.
- Savage, V.M., Webb, C.T., Norberg, J., 2007. A general multi-trait-based framework for studying the effects of biodiversity on ecosystem functioning. *J. Theor. Biol.* 247, 213–229.
- Savage, V.M., Deeds, E.J., Fontana, W., 2008. Sizing up allometric scaling theory. *PLoS Comput. Biol.* 4, e1000171.
- Schmitz, O.J., Buchkowski, R.W., Burghardt, K.T., Donihue, C.M., 2015. Functional traits and trait-mediated interactions: connecting community-level interactions with ecosystem functioning. *Adv. Ecol. Res.* 52, 319–343.
- Shipley, B., 2000. Plasticity in relative growth rate and its components following a change in irradiance. *Plant Cell Environ.* 23, 1207–1216.
- Shipley, B., 2010. *From Plant Traits to Vegetation Structure*. Cambridge University Press, Cambridge, UK.
- Sides, C.B., Enquist, B.J., Ebersole, J.J., Smith, M.N., Henderson, A.N., Sloat, L.L., 2014. Revisiting Darwin’s hypothesis: does greater intraspecific variability increase species’ ecological breadth? *Am. J. Bot.* 101 (1), 56–62.
- Silvertown, J., Poulton, P., Johnston, A.E., Edwards, G., Heard, M., Biss, P.M., 2006. The Park Grass Experiment 1856–2006: its contribution to ecology. *J. Ecol.* 94, 801–814.
- Šimová, I., Violle, C., Kraft, N.J.B., Storch, D., Svenning, J.-C., Boyle, B., Donoghue, J., Jorgensen, P., McGill, B., Morureta-Holme, N., Peet, R.K., Wiser, S.K., Piel, W., Regetz, J., Shildhauer, M., Thiers, B., Enquist, B.J., 2014. Shifts in trait means and variances in North American tree assemblages: species richness patterns are loosely related to the functional space. *Ecography*, in press.
- Sloat, L.L., Henderson, A., Lamanna, C.A., Enquist, B.J., 2014. The effect of the foreshortened drought on carbon exchange in subalpine meadows. *Ecosystems* 18 (3), 533–545.

- Stegen, J.C., Enquist, B.J., Ferrière, R., 2009. Advancing the metabolic theory of biodiversity. *Ecol. Lett.* 12, 1001–1015.
- Stephenson, N.L., Das, A.J., Condit, R., Russo, S.E., Baker, P.J., Beckman, N.G., Coomes, D.A., Lines, E.R., Morris, W.K., Rüger, N., 2014. Rate of tree carbon accumulation increases continuously with tree size. *Nature* 507, 90–93.
- Stevens, R.D., Cox, S.B., Strauss, R.E., Willig, M.R., 2003. Patterns of functional diversity across an extensive environmental gradient: vertebrate consumers, hidden treatments and latitudinal trends. *Ecol. Lett.* 6, 1099–1108.
- Suding, K.N., Goldstein, L.J., 2008. Testing the Holy Grail framework: using functional traits to predict ecosystem change. *New Phytol.* 180, 559–562.
- Suding, K.N., Collins, S.L., Gough, L., Clark, C., Cleland, E.E., Gross, K.L., Milchunas, D.G., Pennings, S., 2005. Functional- and abundance-based mechanisms explain diversity loss due to N fertilization. *Proc. Natl. Acad. Sci. U.S.A.* 102, 4387–4392.
- Suding, K., Lavorel, S., Chapin, F., Cornelissen, J., Diaz, S., Garnier, E., Goldberg, D., Hooper, D., Jackson, S., Navas, M.-L., 2008a. Scaling environmental change through the community-level: a trait-based response-and-effect framework for plants. *Glob. Chang. Biol.* 14, 1125–1140.
- Suding, K.N., Lavorel, S., Chapin, F.S., Cornelissen, J.H.C., Diaz, S., Garnier, E., Goldberg, D., Hooper, D.U., Jackson, S.T., Navas, M.-L., 2008b. Scaling environmental change through the community-level: a trait-based response-and-effect framework for plants. *Glob. Chang. Biol.* 14, 1125–1140.
- Swenson, N.G., 2013. The assembly of tropical tree communities—the advances and shortcomings of phylogenetic and functional trait analyses. *Ecography* 36, 264–276.
- Swenson, N.G., Enquist, B.J., 2007. Ecological and evolutionary determinants of a key plant functional trait: wood density and its community-wide variation across latitude and elevation. *Am. J. Bot.* 94, 451–459.
- Swenson, N.G., Enquist, B.J., 2009. Opposing assembly mechanisms in a Neotropical dry forest: implications for phylogenetic and functional community ecology. *Ecology* 90, 2161–2170.
- Swenson, N.G., Enquist, B.J., Pither, J., Kerkhoff, A.J., Boyle, B., Weiser, M.D., Elser, J.J., Fagan, W.F., Forero-Montaña, J., Fyllas, N., Kraft, N.J.B., Lake, J.K., Moles, A.T., Patiño, S., Phillips, O.L., Price, C.A., Reich, P.B., Quesada, C.A., Stegen, J.C., Valencia, R., Wright, I.J., Wright, S.J., Andelman, S., Jørgensen, P.M., Lacher Jr., T.E., Monteagudo, A., Núñez-Vargas, M.P., Vasquez-Martínez, R., Nolting, K.M., 2012. The biogeography and filtering of woody plant functional diversity in North and South America. *Glob. Ecol. Biogeogr.* 21, 798–808.
- Tilman, D., 1982. *Resource Competition and Community Structure*. (Mpb-17). Princeton University Press.
- Tilman, D., 1999. The ecological consequences of changes in biodiversity: a search for general principles. *Ecology* 80, 1455–1474.
- Tilman, D., 2001. An evolutionary approach to ecosystem functioning. *Proc. Natl. Acad. Sci. U.S.A.* 98, 10979–10980.
- Tilman, D., Lehman, C.L., Thomson, K.T., 1997. Plant diversity and ecosystem productivity: theoretical considerations. *Proc. Natl. Acad. Sci. U.S.A.* 94, 1857–1861.
- Tilman, D., Hillerislambers, J., Harpole, S., et al., 2004. Does metabolic theory apply to community ecology? It's a matter of scale. *Ecology* 85, 1797–1799.
- Vile, D., Shipley, B., Garnier, E., 2006. Ecosystem productivity can be predicted from potential relative growth rate and species abundance. *Ecol. Lett.* 9 (9), 1061–1067.
- Violle, C., Jiang, L., 2009. Towards a trait-based quantification of species niche. *J. Plant Ecol.* 2, 87–93.

- Violle, C., Navas, M.-L., Vile, D., Kazakou, E., Fortunel, C., Hummel, I., Garnier, E., 2007. Let the concept of trait be functional!. *Oikos* 116, 882–892.
- Violle, C., Enquist, B.J., McGill, B.J., Jiang, L., Albert, C.c.H., Hulshof, C., Jung, V., Messier, J., 2012. The return of the variance: intraspecific variability in community ecology. *Trends Ecol. Evol.* 27, 244–252.
- Violle, C., Reich, P., Pacala, S., ENquist, B.J., Kattge, J., 2014. The emergence and promise of functional biogeography. *Proc. Natl. Acad. Sci. U.S.A.* 111 (38), 13690–13696.
- von Allmen, E., Sperry, J., Smith, D., Savage, V., Enquist, B., Reich, P., 2012. A species' specific model of the hydraulic and metabolic allometry of trees II: testing predictions of water use and growth scaling in ring-and diffuse-porous species. *Funct. Ecol.* 26, 1066–1076.
- Wardle, D.A., 2002. Ecology by the numbers. *Trends Ecol. Evol.* 11, 533–534.
- Webb, C.T., Hoeting, J.A., Ames, G.M., Pyne, M.I., Poff, N.L., 2010. A structured and dynamic framework to advance traits-based theory and prediction in ecology. *Ecol. Lett.* 13, 267–283.
- Weiher, E., Keddy, P.A., 1995. Assembly rules, null models, and trait dispersion: new questions from old patterns. *Oikos* 74, 159–164.
- Weiher, E., Freund, D., Bunton, T., Stefanski, A., Lee, T., Bentivenga, S., 2011. Advances, challenges and a developing synthesis of ecological community assembly theory. *Philos. Trans. R. Soc. B Biol. Sci.* 366, 2403–2413.
- West, G.B., Brown, J.H., Enquist, B.J., 1997. A general model for the origin of allometric scaling laws in biology. *Science* 276, 122–126.
- West, G.B., Brown, J.H., Enquist, B.J., 2001. A general model for ontogenetic growth. *Nature* 413, 628–631.
- West, G.B., Woodruff, W.H., Brown, J.H., 2002. Allometric scaling of metabolic rate from molecules and mitochondria to cells and mammals. *Proc. Natl. Acad. Sci. U.S.A.* 99, 2473–2478.
- West, G.B., Enquist, B.J., Brown, J.H., 2009. A general quantitative theory of forest structure and dynamics. *Proc. Natl. Acad. Sci. U.S.A.* 106, 7040–7045.
- Westoby, M., Wright, I.J., 2006. Land-plant ecology on the basis of functional traits. *Trends Ecol. Evol.* 21, 261–268.
- Whittaker, R.H., Levin, S.A., Root, R.B., 1973. Niche, habitat, and ecotope. *Am. Nat.* 107, 321–338.
- Wright, I.J., Reich, P.B., Cornelissen, J.H.C., Falster, D.S., Groom, P.K., Hikosaka, K., Lee, W., Lusk, C.H., Niinemets, U., Oleksyn, J., Osada, N., Poorter, H., Warton, D.I., Westoby, M., 2005. Modulation of leaf economic traits and trait relationships by climate. *Glob. Ecol. Biogeogr.* 14, 411–421.
- Yvon-Durocher, G., Caffrey, J.M., Cescatti, A., Dossena, M., del Giorgio, P., Gasol, J.M., Montoya, J.M., et al., 2012. Reconciling the temperature dependence of respiration across timescales and ecosystem types. *Nature* 487, 472–476.
- Zhang, L., Thygesen, U.H., Knudsen, K., Andersen, K.H., 2013. Trait diversity promotes stability of community dynamics. *Theor. Ecol.* 6, 57–69.
- Zhao, M., Running, S.W., 2010. Drought-induced reduction in global terrestrial net primary production from 2000 through 2009. *Science* 329, 940–943.
- Zhao, M., Heinsch, F.A., Nemani, R.R., Running, S.W., 2005. Improvements of the MODIS terrestrial gross and net primary production global data set. *Remote Sens. Environ.* 95, 164–176.